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TITLE OF INVENTION RECOMBINANT CELL LINE AND SCREENING METHOD FOR IDENTIFYING AGENTS
WHICH REGULATE APOPTOSIS AND TUMOR SUPPRESSION

APPLICANT(S) FOR DO/EO/US

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Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1).
4. ☒ A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
5. ☐ A copy of the International Application as filed (35 U.S.C. 371(c)(2))
 - a. ☐ is transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☐ has been transmitted by the International Bureau.
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US)
6. ☐ A translation of the International Application into English (35 U.S.C. 371(c)(2)).
7. ☐ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3))
 - a. ☐ are transmitted herewith (required only if not transmitted by the International Bureau).
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 - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
 - d. ☐ have not been made and will not be made.
8. ☐ A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. ☐ An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)).
10. ☐ A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).

Items 11. to 16. below concern other document(s) or information included:

11. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
12. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
13. ☐ A **FIRST** preliminary amendment.
☐ A **SECOND** or **SUBSEQUENT** preliminary amendment.
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15. ☐ A change of power of attorney and/or address letter.
16. ☒ Other items or information:

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RECOMBINANT CELL LINE AND SCREENING METHOD
FOR IDENTIFYING AGENTS WHICH REGULATE APOPTOSIS AND
TUMOR SUPPRESSION

09/674876

CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims priority under 35 U.S.C 119(e) from U.S. Provisional Application Nos. 60/084,664 filed May 7, 1998, 60/091,391, filed July 1, 1998, 60/092,871 filed July 15, 1998 and 60/107,689 filed November 9, 1998, the disclosure of each being incorporated by reference herein.

Pursuant to 35 U.S.C. §202(c) it is acknowledged that the U.S. Government has certain rights in the invention described herein, which was made in part with funds from the National Institutes of Health, Grant Number CA-60088.

FIELD OF THE INVENTION

The present invention relates to the identification of anti-neoplastic agents. Specifically, the invention provides novel cell-based assays and reagents for screening compounds involved in the regulation of apoptosis and tumor suppression.

BACKGROUND OF THE INVENTION

Several publications are referenced in this application by author name and year of publication in parentheses in order to more fully describe the state of the art to which this invention pertains. Full citations for these references are found at the end of the specification. The disclosure of each of these

publications is incorporated by reference herein.

Apoptosis is a genetically controlled pathway of programmed cell death. The hallmarks of apoptosis are cytoplasmic boiling, chromatin condensation, and DNA fragmentation. Apoptosis plays a major role in development and disease. Regulation of cell death is essential for normal development and is an important defense against viral infection and the emergence of cancer. Excessive cell death can lead to impaired development and degenerative diseases, whereas too little cell death can lead to cancer and persistent and sustained viral infection.

Tumor suppressor genes are a class of genes involved in different aspects of normal control of cellular growth and division. Inactivation of tumor suppressor genes, usually by genetic means, contributes to tumor development. The first evidence for the existence of such genes came from the observation that fusion of normal cells with tumor cells resulted in hybrids which were nontumorigenic. However, with the loss of chromosomes, which occurs naturally when hybrid cells are grown in culture, the tumorigenic phenotype was sometimes restored. Furthermore, the introduction of normal chromosomes into tumorigenic hybrids can cause reversal to the nontumorigenic phenotype. Thus expression of genes from the normal parent suppresses tumorigenicity.

As the normal functioning of tumor suppressor genes suppresses tumorigenicity, regions of chromosomes which are lost in tumors are likely to carry tumor suppressor genes. Karyotype analysis of a wide variety of tumor types has shown that certain chromosomal deletions occur with a high frequency and in a tumor specific manner,

thus implicating the chromosomal region as a likely location of tumor suppressor genes.

Apoptosis is controlled through the expression of an increasing number of genes conserved in nematodes through mammals and viruses. Some gene products are activators of apoptosis, whereas others are inhibitors. Tumor suppressor genes are also widely conserved. The characterization of the function of these gene products will facilitate elucidation of the molecular mechanisms which regulate cell growth at the biochemical level. This knowledge in turn, facilitates the identification and characterization of agents suitable for regulating apoptotic cell death and tumor suppression which may be used to advantage in the treatment of malignancy and other disorders characterized by aberrant cell growth.

SUMMARY OF THE INVENTION

In accordance with the present invention, it has been discovered that inhibition of the transcriptional coactivator p300 induces p53 accumulation resulting in apoptotic cell death. p300 is required to transactivate the p53 inhibitor mdm2. Failure of p300 to transactivate mdm2 results in accumulation of p53 to high levels causing apoptosis, a desirable characteristic of anti-cancer therapeutics. The present invention is directed to the identification of p300 transactivating inhibitors which act to induce p53 accumulation through the failure to coactivate mdm2. Such agents should be effective to induce apoptotic cell death for the treatment of proliferative disorders associated with aberrant cell growth.

The present invention provides novel cell lines and methods for identifying agents that regulate apoptosis.

According to one aspect of the invention a recombinant cell line is provided for identifying and characterizing p300 inhibitors which regulate apoptosis. An exemplary cell line of the invention comprises a) a first plasmid expressing a p300 responsive promoter operably linked to a first reporter gene; b) a second plasmid expressing a non p300 responsive promoter operably linked to a second reporter gene; and c) a third plasmid expressing a selectable marker gene. The cell line may optionally include a plasmid encoding wild -type p300 to augment endogenously expressed p300 protein levels.

In another embodiment of the invention, a screening method is provided using the cell line of the invention to identify therapeutic agents that regulate apoptosis. The screening method entails determining whether a potential therapeutic reagent inhibits p300 activity thereby inducing apoptosis. One such method is performed by contacting the recombinant cells of the invention with the therapeutic agent. The recombinant cells include a first plasmid expressing a p300 responsive promoter operably linked to a first reporter gene, a second plasmid expressing a non p300 responsive promoter operably linked to a second reporter gene; and a third plasmid expressing a selectable marker gene. Following administration of the reagent repression of the p300 responsive reporter and repression of the non-p300 responsive reporter gene by the therapeutic reagent would be assessed via the levels of reporter gene activity. Repression of the p300 responsive reporter gene and not the non-responsive p300 reporter gene indicates that the compound inhibits p300 transactivation and thereby may be efficacious in inducing apoptosis in target cells. In an alternative

embodiment, plasmids may be constructed which contain a plurality of the above described gene constructs.

In yet another embodiment of the invention, a Bcl-2 associated transcription factor (referred to herein as BP-1 and btf) having the sequence of SEQ ID NO: 1 has been identified which interacts with ElB 19K as well as with anti-apoptotic family members, Bcl-2 and Bcl-x_L, but not the proapoptotic protein, Bax. Btf protein has a sequence of SEQ ID NO:2, binds DNA in vitro and represses transcription in reporter assays. The Btf gene has been localized to chromosome 6 (6q22-23), a region that is deleted in certain cancers providing support for Btf's role as a tumor suppressor. Btf is also active in a unique pathway by which Bcl-2 regulates apoptosis.

The availability of the DNA sequence encoding btf facilitates the production of large amounts of btf protein in suitable expression systems. The Btf protein so generated may be used to advantage in the production of antibodies or antibody fragments that may be used to detect Btf protein in clinical samples.

Elucidation of the interaction between Bcl-2 and Btf facilitates the identification and characterization of novel therapeutic agents that specifically disrupt the binding the Bcl-2 and Btf, while not affecting Bcl-2 binding to other proteins. Agents that disrupt binding between Bcl-2 and Btf may be assessed in cell lines which co-express the two proteins. Such agents may comprise peptides or other small molecule inhibitors which act competitively to inhibit the binding between Bcl-2 and Btf, there by effectuating cell death in a variety of tumors.

In accordance with the present invention, methods

are provided for screening tumor samples for deletion of the Btf gene and/or protein. Such methods may be either immunological and employ Btf specific antibodies to detect the presence or absence of btf protein expression. Alternatively, the methods may be genetically based. Btf specific oligonucleotides may be employed to assess the sample for the presence or absence of btf encoding nucleic acids. In patients having a btf deletion, the Btf gene may be reintroduced via genetic engineering methods.

The cell lines and screening methods of the present invention provide a unique and advantageous testing system for evaluating reagents that regulate apoptosis. Such agents may then be used to advantage to treat pathological conditions related to aberrant cell growth, including cancer.

BRIEF DESCRIPTION OF THE DRAWINGS

Figures 1A and 1B show the results of Western blot analysis of p53, Mdm2, Bax, and p21^{WAF1} in p53A and LTR.1A cells after incubation at the nonpermissive (37.5°C) and permissive (32°C) temperature at day 0 (lanes 1 and 6), day 1 (lanes 2 and 7), day 2 (lanes 3 and 8), day 3 (lanes 4 and 9) and day 5 (lanes 5 and 10).

Figure 2 shows the results obtained from indirect immunofluorescence of Mdm2 in p53A (top panel) and LTR.1A (bottom panel) cells after incubation at 32°C from day 0 to day 3.

Figures 3A and 3B are a pair of graphs showing cell viability as demonstrated by trypan blue staining

of p53A and LTR.1A cells at the nonpermissive (37.5°C) and permissive (32°C) temperature from day 0 to day 3.

Figure 4 depicts a series of micrographs showing light microscopy analysis of p53A and LTR.1A cells incubated at 32°C from day 0 to day 3 to detect apoptotic cells.

Figures 5A and 5B show the results of Northern blot analysis of *mdm2* expression of p53A cells incubated at 32°C for the indicated times. Fig. 5A, autoradiograph of blot; Fig. 5A ethidium stained gel showing loading and integrity of RNA.

Figure 6 is an autoradiograph showing the results of Western blot analysis of ectopically expressed HA-tagged p300, endogenous Mdm2, and actin in p53A cells transiently transfected by electroporation with pCMVβ alone (lane 1 - 12 μg and lane 2 - 24 μg), pCMVβp300 (p300 wild-type) (lane 3 - 12 μg and lane 4 - 24 μg) or p300Δ30 (p300 mutant) (lane 5 - 12 μg and lane 6 - 24 μg).

Figure 7 shows the results of indirect immunofluorescence of HA-tagged p300 and endogenous Mdm2 in p53A cells transiently transfected with 24 μg of vector alone, wild-type p300 and mutant p300 (arrows indicate cells with ectopic p300 expression and up-regulation of Mdm2).

Figure 8 is a bar graph showing cell viability, as measured by trypan blue staining, of p53A cells

transiently transfected with vector alone, wild-type p300 or mutant p300.

5 Figure 9 is a graph showing the percentage of apoptotic cells that are positive for ectopic expression of a control lamin fragment, p300 wild-type and p300 mutant determined by cell morphology using indirect immunofluorescence in p53A cells transiently transfected with 24 μ g of pCEP4-LA(1-406). pCMV β p300 or pCMV β p300 Δ 30
10 plasmid DNAs.

Figure 10 shows the results of Western Blot analysis of E1A, Mdm2 and actin in LTR.1A cells transiently transfected with 10 μ g of vector alone (lane
15 1), 12S E1A (lane 2), 12SRG2 (lane 3), or 12S.YH47.928 (lane 4).

Figure 11 shows cell viability, as measured by trypan blue exclusion, of LTR.1A cells transiently transfected with vector alone, 12S E1A, 12S.RG2 or
20 12S.YH47.928.

Figure 12 shows Western blot analysis of p53A cells transiently transfected with 5 and 10 μ g of pCMV vector alone, *mdm2* wild-type (pCOC-X₂) or *mdm2* mutant (pCOC- Δ XM)
25 plasmid DNAs.

Figure 13 is a graph showing cell viability as assessed by trypan blue staining of p53A cells transiently transfected with *mdm2* expression plasmids
30 incubated at 32°.

Figure 14 is a graph showing the percentage of

apoptotic cells that were positive for ectopic expression of a control lamin fragment, Mdm2 wild-type, and Mdm2 mutant determined by cell morphology using indirect immunofluorescence in p53A cells transiently transfected with 10 μ g of pCEP4-LA(1-406), pCOC-X₂ (wild-type Mdm2) or pCOC- Δ XM (truncated Mdm2) plasmid DNAs.

Figures 15A, 15B and 15C show the results of Western blot analysis of p53, Mdm2, bax, and p21Waf1 in p53A cells stably expressing PSVneo (p53An1) E1B19K (19K1) or Bcl-2 (4B).

Figure 16 shows Western blot analysis of Mdm2, Bcl-2 E1B 19K and actin in p53An1 cells transfected transiently in pCND43 alone, (lane 1); pCMV19K (lane 2) and pCNA3myc-tagged bcl-2, (lane 3).

Figure 17 shows the results of Northern blot analysis of Mdm2 expression using 30 μ g of cytoplasmic RNA of 19K1 cells incubated at 32°C for the indicated time intervals.

Figure 18 is a schematic diagram illustrating the regulation of cell death in p53A, LTR.1A and 19K1/4B cells.

Figures 19A and 19B are growth profiles of transformed yeast cells demonstrating that BP-1 interacts with E1B 19K in yeast. The yeast two-hybrid assay was utilized to demonstrate binding between BP-1 and E1B 19K. Growth in the presence of histidine indicates that both plasmids can be expressed in yeast, and growth in the absence of histidine demonstrates an

interaction between the two proteins. Apc-2 represents an irrelevant hydrophobic protein used as a negative control. The specificity of the interaction between BP-1 and E1B 19K was tested using related proteins (Bcl-2, Bcl-x_L, and Bax) as well as missense mutants of E1B 19K (pm7, pm51, pm87, and pm102, Fig. 19A. Fig. 19B shows growth profiles which reveal the minimal regions of E1B 19K required for interaction with BP-1 were mapped using a series of deletion mutants. Figure 19C is a schematic representation of E1B 19K showing the missense and deletion mutants tested in the two-hybrid assay. Regions I and III indicate the locations of the Bcl-2 homology domains, BH1 and BH3. E1B 19K does not contain recognizable BH2 or BH4 domains.

Figures 20A and 20B are autoradiographs showing the results of Northern blot analysis performed to determine *btf* expression in various tissues (Fig. 20A), and cancer cell lines (Fig. 20B). Two transcripts were observed at 5 kB (*btf_L*) and 3 kB (*btf_s*). These transcripts appeared to be ubiquitously expressed, except that *btf* was not detected in Raji cells derived from Burkitt's lymphoma. Lower panels depict β -actin expression to assess the quality of the RNA and to control for loading efficiency.

Figure 21 depicts the amino acid sequence of Btf. Btf_L, identified in the GenBank database (accession # D79986) is predicted to encode a 918 amino acid protein shown here. Btf_s, which was obtained in the λ -screen, contains the complete Btf_L sequence except that it is missing 49 amino acids between residues 797 and 846 (bold) present in Btf_L and BP-1. The underlined regions

represent the positions of nuclear localization sequences.

Figure 22 is a schematic representation of the Btf variants and deletion mutants used to characterize Btf function. The gray and black boxes represent the locations of putative bZIP and Myb-DNA binding domains, respectively. BP-1 (N384), identified in the yeast two-hybrid screen for E1B 19K binding proteins, contains residues 384-918 of Btf_L. Deletion mutants of Btf_S, N552 and C210 were used in transcriptional reporter assays to determine the regions of Btf_S that contribute to transcriptional repression.

Figure 23 is an autoradiograph showing ³⁵[S]-methionine labeled *in vitro* translated Myc-Btf_S and E1B 19K prepared using the TNT T7 reticulocyte lysate system and incubated with native DNA-cellulose in NETN buffer for 2 hrs. The data show that Btf_S binds to DNA-cellulose *in vitro*. Samples were washed five times in NETN and the proteins were resolved by 17% SDS-PAGE. Immunoprecipitated proteins (lanes 1 and 2) were analyzed to confirm the presence of the *in vitro* translated products.

Figure 24 is an autoradiograph showing *in vitro* interactions between Btf_S and anti-apoptotic Bcl-2 family members. ³⁵[S]-labeled *in vitro* translated Myc-Btf_S or Myc-Bax was combined with V5/His-E1B 19K, Bcl-2, Flag-Bcl-x_L, or luciferase prepared using the TNT T7 reticulocyte lysate system. The proteins were immunoprecipitated with anti-Myc monoclonal antibody in NETN buffer for 1.5 hrs followed by protein A-sepharose

for 0.5 hr. Samples were washed in NETN buffer,
resolved by 14% SDS-PAGE and visualized by
autoradiography. In addition, 1 μ l of each of
translation reaction was analyzed to verify that equal
5 amounts of proteins were used in the binding assay.

Figure 25 is a series of micrographs showing that
the nuclear subcellular localization of Btf₃ is altered
by E1B 19K, Bcl-2 and Bcl-x_L. HeLa cells were
10 transfected with expression plasmids pcDNA3-Myc- Btf₃
alone and with pCMV E1B 19K or pcDNA3-Bcl-2 as
indicated. Cells were fixed 24 hours post-transfection
and double stained against Myc and E1B 19K or Bcl-2. All
cells were photographed with an original magnification
15 of 1000X.

Figure 26 is a graph showing the percent of nuclear
Btf₃ expression (Btf₃ /total Btf₃) in HeLa cells
expressing Bcl-x_L transfected with pcDNA3-Myc-Btf₃ and
20 stained 24 hours post-transfection against Myc.

Figure 27 is a graph showing the results of a
reporter assay demonstrating that Btf₃ is a transcription
repressor and is inhibited by Bcl-2-like proteins. HeLa
25 cells were transfected with 2.5 μ g of the reporter
construct (luciferase construct containing GAL-4 DNA-
binding sites within its promoter), 2.5 μ g of GAL-4 DNA-
binding domain fusion genes (pml- Btf₃, Δ -N552, and Δ -C210
or empty pml vector control), and 2.5 μ g of bcl-2 family
30 gene (pCMV-E1B 19K, pcDNA3-Bcl-2, or pcDNA3-Flag-Bcl-x_L,
or empty pcDNA3 vector control). Cells were harvested
24 hours post-transfection and the luciferase activity
was measured in a scintillation counter using the

luciferase substrate, luciferin. Values were normalized for protein concentrations measured by a Bradford assay and graphed as a percent of the negative control (empty pml vector). The experiment was repeated six times and the high and low values for each sample were dropped. Bars indicate standard deviation, n=4.

Figures 28A and 28B are a pair of graphs showing that Btf₂ inhibits transformation by E1A and E1B 19K (Fig. 28A) or p53DD (Fig. 28B). Primary baby rat kidney (BRK) cells were transfected with carrier DNA along with linearized test DNA (15 µg pCMV-E1A, 15 µg pCMV-E1B 19K or pCMV-p53DD, and 45 µg pcDNA3-Myc-Btf₂). DNA concentrations were kept constant using appropriate empty vectors. The cells were cultured for 3 to 4 weeks and then stained with Giemsa. Foci were counted from four dishes per condition. Bars indicate standard deviation, n=4.

Figure 29 shows a representative FACS scan from Table 1 at 72 hours post-transfection of pIRES-EGFP-Btf₂ and the empty pIRES-EGFP vector into HeLa cells. The data reveal that Btf₂-mediated cell death occurs by apoptosis. Cell cycle kinetics from propidium iodide staining are shown for the total cell population as well as the EGFP-positive cells.

Figure 30 is a series of micrographs showing transfected cells incubated with Hoechst dye 72 hours post-transfection to visualize the DNA. Cells transfected with pIRES-EGFP-Btf₂ (right side) were compared to control pIRES-EGFP (left side). Arrows for each set correspond to the same cell visualized with

brightfield (top), EGFP (green; middle), and Hoechst dye (blue; bottom). The presence of condensed chromatin in the presence of Btf₃ indicates cell death by apoptosis. The cells were photographed with an original magnification of 1000X.

DETAILED DESCRIPTION OF THE INVENTION

The present invention provides cell lines and screening assays and reagents for identifying and characterizing compounds which regulate apoptosis.

The tumor suppressor p53 gene product is a negative regulator of cellular growth and transformation (Ko and Prives 1996; Levine 1997; Vogelstein and Kinzler 1992; White 1996). Many studies have demonstrated that p53 functions as a transcriptional regulator by sequence-specific DNA binding (El-Deiry et al. 1992; Funk et al. 1992; Pietsenpol et al. 1994). The p53 gene product regulates transcription by activation or repression (Ko and Prives 1996). DNA damage increases p53 levels which promotes cell cycle arrest allowing DNA repair to occur. Alternatively DNA damage can induce apoptosis presumably when such damage is beyond repair (Ko and Prives 1996; Levine 1997; White 1996). The growth arrest function of p53 is predominantly implemented by transactivation of the cyclin dependent kinase inhibitor *p21/WAF1/CIP1* (El-Deiry et al. 1993). p53 can also induce apoptosis by up-regulating the death-promoting *bax* gene (Miyashita and Reed 1995). Other transcriptional targets for p53 include *GADD45* (Kastan et al. 1992), *mdm2* (Barak et al. 1993; Wu et al. 1993), *cyclin G* (Okamoto and Beach 1994), and *IGF-BP3* (Buckbinder et al. 1995). The DNA binding ability of p53 plays a major role in regulating cell growth. The most frequently occurring p53

mutations in human tumors are found in this DNA binding domain (Hollstein et al. 1991; Ko and Prives 1996). Accordingly, intact p53 transcriptional functions are important to maintain genomic integrity.

5 The *mdm2* gene was originally cloned because of its amplification in a spontaneously transformed murine BALB/c cell line (Fakharzadeh et al. 1991). The human homologue of Mdm2 protein is a negative regulator of p53. Mdm2 protein inhibits p53-mediated functions of G₁ arrest and apoptosis (Chen et al. 1996), most likely by binding to the N-terminal transactivation domain of p53 (Momand et al. 1992; Oliner et al. 1993). Furthermore, Mdm2 appears to direct p53 degradation via the ubiquitin pathway (Haupt et al. 1997; Kubbutat et al. 1997). The *mdm2* promoter contains p53 binding consensus sequences where p53 binds and positively regulates its expression, creating a negative feedback loop for regulating the activity and levels of p53 (Barak et al. 1993; Haupt et al. 1996; Wu et al. 1993). The functional interdependence of Mdm2 and p53 was exemplified in studies with knock-out mice. Loss of Mdm2 resulted in early embryonic lethality which was rescued by deletion of p53 (Donehower et al. 1992; Montes de Oca Luna et al. 1995). Thus, Mdm2 is required *in vivo* for down-modulation of p53 function and perturbation of this regulation can be deleterious to embryonic development.

 The CBP/p300 family members regulate transcription by functioning as transcriptional coactivators. Although the precise mechanism of how transcriptional adaptors function is not known, CBP/p300 and an interacting protein, P/CAF, have been shown to have histone acetyltransferase activity (Bannister and Kouzarides 1996; Ogryzko et al. 1996; Yang et al. 1996),

implicating a role for histone acetylation in transcriptional regulation. These proteins also interact with several transcription factors such as the TAFs (Thut et al. 1995), TBP (Abraham et al. 1993), CREB (Chrivia et al. 1993; Kwok et al. 1994), c-Jun/v-Jun (Bannister and Kouzarides 1995), c-Myb/v-Myb (Dai et al. 1996), c-Fos (Bannister and Kouzarides 1995) and others, which may determine the specificity of the regulation. The p300 family of proteins has been recently shown to bind to p53 and function as coactivators of p53-inducible genes (Avantaggiati et al. 1997; Gu et al. 1997; Lill et al. 1997; Scolnick et al. 1997). The N-terminal activation domain of p53 directly interacts with the carboxy-terminal of p300 (Gu and Roeder 1997). It has also been shown that p300 can acetylate the C-terminal domain of p53 and that this modification increases the sequence-specific DNA binding ability of p53 (Gu and Roeder 1997). Thus, acetylation of specific transcription factors may reflect one level of p300 transcriptional regulation.

The adenoviral early region 1 (E1) genes encode for proteins that aid in cellular transformation by activating proliferation and suppressing apoptosis (White 1993; White and Gooding 1994). Expression of the adenoviral E1A gene stimulates cell cycle progression by interacting with and subverting the function of cellular proteins required for normal cell cycle and transcription regulation. E1A interacts with the retinoblastoma (Rb) gene product as well as its family members, p107 and p130 (Dyson and Harlow 1992; Moran 1993; Whyte et al. 1988). E1A also binds to and sequesters p300 (Eckner et al. 1994; Moran 1993; Yang et al. 1996). E1A interactions with these cellular

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proteins are important for transformation as suggested by the observation that E1A mutants that fail to interact with these proteins are incapable of promoting transformation. Expression of E1A alone, however, is insufficient to transform primary baby rat kidney (BRK) cells because cell cycle deregulation by E1A also stimulates p53-dependent apoptosis. Binding of p300 to E1A cosegregates with induction of p53 and stabilization (Chiou and White 1997; Querido et al. 1997; Sanchez-Prieto et al. 1995). Transcriptional activation of p53 target genes, such as *bax* (Han et al. 1996; Sabbatini et al. 1995), followed by induction of caspase activation implements cell death (Rao and White 1997; Sabbatini et al. 1997). Transcriptional activation of the p53 target gene *p21^{WAF1}* is also induced, which contributes to implementation of cell cycle arrest by p53, which is not apparent due to cell death (Han et al. 1996; Sabbatini et al. 1995).

E1A-induced cellular transformation is sustained by coexpression of the Bcl-2 adenoviral homologue, E1B 19K or Bcl-2 itself, which inhibit E1A-induced, p53-mediated apoptosis. E1B 19K and Bcl-2 inhibit apoptosis in part by binding to the death promoting Bax protein (Han et al. 1996). Thus E1B 19K or Bcl-2 expression inhibits p53-mediated apoptosis but not growth arrest (Chiou et al. 1994; Debbas and White 1993; Han et al. 1996; Sabbatini et al. 1995). These cooperative functions between early adenoviral genes stimulating cell cycle progression and inhibiting apoptosis ensures efficient transformation.

In accordance with the present invention it has been discovered that p300 is specifically required for transactivation of the *mdm2* gene by p53 and also for

regulating p53-mediated apoptosis. Cells expressing E1A were unable to up-regulate Mdm2, causing p53 to be induced to high levels which resulted in p53-dependent apoptosis. In contrast, BRK cells expressing c-Myc instead of E1A, upregulated Mdm2, causing p53 down-regulation and inhibition of p53-dependent apoptosis. Thus, the inability to down-regulate p53 is associated with apoptosis and provides an explanation for the differences between the p53-dependent apoptotic response of E1A versus Myc transformed cells.

The adenovirus E1B 19K protein is a potent inhibitor of apoptosis and cooperates with E1A to transform primary rodent cells. E1B 19K shows sequence and functional homology to the mammalian anti-apoptotic gene product, Bcl-2. Like Bcl-2, the biochemical mechanism of E1B 19K function includes binding to and antagonizing cellular pro-apoptotic proteins, such as Bax, Bak, and Nbk/Bik. In addition, there is evidence that E1B 19K can affect gene expression, but whether this contributes to the anti-apoptotic function of 19K has not been determined. In an effort to further understand the functions of E1B 19K, we screened for 19K-associated proteins using the yeast two-hybrid system. A novel protein Btf (Bcl-2-associated transcription factor) was identified that interacts with E1B 19K, as well as with anti-apoptotic family members, Bcl-2 and Bcl-xL, but not the pro-apoptotic protein Bax. btf is a widely expressed gene that encodes a protein with homology to the basic zipper (bZip)- and Myb-DNA binding domains. Btf binds DNA *in vitro* and represses transcription in reporter assays. E1B 19K, Bcl-2, and Bcl-xL sequester Btf in the cytoplasm and block its transcriptional repression activity. Expression of Btf

also inhibited transformation by E1A with either E1B 19K or mutant p53, suggesting a role in either promoting apoptosis or cell cycle arrest. Indeed, the sustained over-expression of Btf in HeLa cells induced apoptosis which was inhibited by E1B 19K. Furthermore, the chromosomal localization of *btf* (6q22-23) maps to a region that is deleted in some cancers, consistent with a role for Btf in tumor suppression. Thus, *btf* may represent a novel tumor suppressor gene, residing in a unique pathway by which the Bcl-2 family can regulate apoptosis.

These observations have facilitated the design of recombinant cell lines that may be used to advantage to screen therapeutic agents that regulate apoptosis.

The nucleic acids of the invention are sometimes referred to herein as "isolated nucleic acids". This term, when applied to DNA, is intended to signify a DNA molecule that is separated from sequences with which it is immediately contiguous (in the 5' to 3' directions) in the naturally occurring genome of the organism from which it was derived. For example, the "isolated nucleic acids" of the invention may comprise a DNA molecule inserted into a vector, such as a plasmid or a virus vector, or integrated into the genomic DNA of a procaryote or eucaryote. With respect to RNA molecules of the invention, the term "isolated nucleic acids" primarily refers to RNA molecules encoded by isolated DNA molecules as defined above, or produced by synthetic methods.

The term "transcription control (or regulatory) element" as used herein, refers to an isolated DNA segment that, under specified conditions, possesses a transcription-regulating activity with respect to the

expression of a transcription unit (i.e., a gene, as defined below).

The term "enhancer" is used herein to refer to a type of transcription control element. Enhancers
5 generally activate the expression of a cis-linked gene.

The terms "transcription control element," "transcription regulatory element" and "enhancer" are used interchangeably herein when referring to the isolated nucleic acid molecules of the present
10 invention.

The term "transcription factor" refers to protein factors that interact with DNA to enhance or repress transcription.

The term "repressor" refers to a transcription
15 factor that inhibits gene expression upon interaction with DNA.

The term "transcription unit" refers to a nucleic acid molecule comprising one or more sequences which encode a gene product (usually a protein) and which is
20 operably linked to a promoter or other regulatory sequences (enhancers etc.) necessary for expression of the coding sequence. The term "gene" is often used interchangeably with the term "transcription unit".

The term "operably linked" means that the
25 regulatory sequences necessary for expression of the coding sequence are placed in the DNA molecule in the appropriate positions relative to the coding sequence so as to effect expression of the coding sequence. This same definition is sometimes applied to the arrangement
30 of transcription units and other transcription control elements (e.g. enhancers) in an expression vector.

The term "reporter gene" refers to a gene whose expression may be assayed; such genes include, without

limitation, *lacZ*, amino acid biosynthetic genes, e.g., the yeast *LEU2*, *HIS3*, *LYS2*, or *URA3* genes, nucleic acid biosynthetic genes, the mammalian chloramphenicol transacetylase (CAT) gene, the luciferase gene, the green fluorescent protein (GFP) or any surface antigen gene for which specific antibodies are available.

A "promoter" is a DNA sequence located proximal to the start of transcription at the 5' end of an operably linked transcribed sequence. The promoter may contain one or more regulatory elements or modules which interact in modulating transcription of the operably linked gene.

The term "tumor suppressor", as used herein, relates to any protein/polypeptide inhibiting growth of tumor cells in-vitro and/or in-vivo. Growth inhibition involves mechanisms such as control of apoptosis and/or of cell cycle progression as well as mechanisms unidentified so far. "Tumor suppressors" are proteins displaying biological activities identical to or similar to those of p53, Rb (retinoblastoma gene product), WT (Wilms tumor suppressor gene), VHL (von Hippel-Lindau tumor suppressor gene), BRCA1 (breast cancer susceptibility gene) and p16 (cyclin-dependent kinase inhibitor).

I. METHODS FOR CONSTRUCTING PLASMIDS AND PRODUCING THE CELL LINES OF THE INVENTION

The cell lines of the invention may be prepared according to the general methods set forth below for plasmid construction, transformation of cultured cells, and maintenance of cell lines.

A. Plasmids

The plasmids of the invention may be constructed using conventional cloning methods. A variety of different plasmid vectors are commercially available. Protocols for constructing such plasmids are provided in Current Protocols in Molecular Biology, Ausubel et al. (editors), John Wiley & Sons, Inc. 1987-1997. Detailed protocols for constructing the plasmids of the invention are provided hereinbelow.

B. Cell Lines

To achieve stable gene transfer, plasmid DNA must first be introduced into host cells. This may be accomplished according to numerous methods known in the art, including but not limited to 1) calcium phosphate precipitation; 2) electroporation; and 3) liposome-mediated transfection. For general protocols, see, e.g., Chapter 9 in Current Protocols in Molecular Biology, Ausubel et al. (editors), John Wiley & Sons, Inc. 1987-1997.

In accordance with the present invention, cell lines can be generated by co-transfection of a plurality of plasmids. Alternatively, a sequential transfection can be performed, whereby a first cell line is generated that has stably incorporated a plasmid containing a p300 regulated promoter, such as the mdm2 promoter, operably linked to a reporter gene such as CAT. This cell line can then be transfected in turn with a plasmid encoding wild type p300 to augment endogenous p300 levels.

Stable transfectants are selected by the ability of an individual cell colony to grow in the presence of a selected antibiotic, by virtue of a resistance-encoding gene carried on the transfecting plasmid DNA and

incorporated into the genome of the cell. In a preferred embodiment of the invention, both of the plasmids used would contain a selectable marker gene for double selection of stable transfectants by antibiotic resistance.

II. USES OF CELL LINES FOR SCREENING AGENTS THAT REGULATE APOPTOSIS

The recombinant cell lines of the invention may be used to identify and characterize therapeutic compounds that act as p300 inhibitors and thereby induce apoptosis.

For example, cells may be transfected with a plasmid containing a reporter gene operably linked to a promoter which is transactivated by p300. An exemplary promoter is the mdm2 promoter. The cells would also be co-transfected with a control plasmid containing a second reporter gene operably linked to a non-p-300 responsive promoter. The cell line may optionally be stably transfected with a cDNA encoding p300 to augment endogenous p300 levels. Such cell lines may be used in reporter assays for compounds that inhibit p300-dependent transcription. These cells lines can be screened for repression of the mdm-2 linked reporter by compounds that do not alter expression of the non-p300 responsive reporter gene.

Another strategy entails assaying p300 directly for molecules that bind to the E1A binding site on p300. Since binding of E1A to p300 inhibits p300 function, any molecule or compound that mimicked E1A, should be a potential p300 inhibitor. An E1A mutant that fails to

bind p300 may be used as a control in such an assay.

Additionally, matched cell lines derived from the LTR 1A (myc +tsp53[val135]) cell line, exemplified herein, can be used in an in vivo cell-based screen for p300 inhibitors which, like E1A, promote p53 stabilization by inhibiting mdm-2 transcription and cause apoptosis. LTR 1A cells have intact p300 function and the ts p53 at the permissive temperature retains the capacity to activate the endogenous mdm-2 gene which is required for cell survival. Potentially therapeutic compounds that inhibit p300 function would cause apoptosis at the permissive temperature. This apoptotic phenotype can be used to advantage to identify agents of interest. LTR 1A cells that ectopically express Mdm-2 from a heterologous promoter would not be susceptible to p300 inhibition and are used as a control for p300 specificity.

The availability of the DNA sequences encoding Btf and Bcl-2 also facilitates the design of additional cell lines and screening methods to identify agents which regulate apoptosis via this pathway. Btf and Bcl-2 may be overexpressed and agents that disrupt Btf/Bcl-2 binding assessed. Disruption of Btf/Bcl-2 binding may be assessed using co-immunoprecipitation for example. Agents so identified are expected to induce apoptosis in the targeted cell. Detailed methods for detecting apoptosis in cells are provided hereinbelow.

Antibodies specific for Btf are may be used to assess tumor samples for the presence or absence of Btf protein. Methods for generating antibodies and antibody fragments are well known to those of ordinary skill in the art. Screening methods employing such antibodies

include but are not limited to Western blotting, immunofluorescence and affinity chromatography.

Oligonucleotide probes and primers may also be used to assess expression of Btf in tumor samples in the clinical setting. DNA would be isolated from the tumor sample in question and assessed for the presence or absence of btf encoding nucleic acids. In accordance with the present invention, nucleic acids having the appropriate level of sequence homology with Sequence I.D. No. 1 may be identified by using hybridization and washing conditions of appropriate stringency. For example, hybridizations may be performed, according to the method of Sambrook et al., (supra) using a hybridization solution comprising: 5X SSC, 5X Denhardt's reagent, 1.0% SDS, 100 µg/ml denatured, fragmented salmon sperm DNA, 0.05% sodium pyrophosphate and up to 50% formamide. Hybridization is carried out at 37-42°C for at least six hours. Following hybridization, filters are washed as follows: (1) 5 minutes at room temperature in 2X SSC and 1% SDS; (2) 15 minutes at room temperature in 2X SSC and 0.1% SDS; (3) 30 minutes-1 hour at 37°C in 1X SSC and 1% SDS; (4) 2 hours at 42-65°C in 1X SSC and 1% SDS, changing the solution every 30 minutes.

One common formula for calculating the stringency conditions required to achieve hybridization between nucleic acid molecules of a specified sequence homology is (Sambrook et al., 1989):

$$T_m = 81.5^{\circ}\text{C} + 16.6\text{Log} [\text{Na}^+] + 0.41(\% \text{G+C}) - 0.63 (\% \text{formamide}) - 600/\text{\#bp in duplex}$$

As an illustration of the above formula, using $[\text{Na}^+] = [0.368]$ and 50% formamide, with GC content of 42% and an average probe size of 200 bases, the T_m is

57°C. The T_m of a DNA duplex decreases by 1 - 1.5°C with every 1% decrease in homology. Thus, targets with greater than about 75% sequence identity would be observed using a hybridization temperature of 42°C.

5 If a tumor cell is found to contain a Btf deletion, the tumor cells may be genetically engineered to express the btf gene. Methods for delivering nucleic acids to target cells include retroviral targeting vectors and adenoviral vectors.

10 The following materials and methods are provided to facilitate the practice of the present invention.

15 *Cells and culture conditions*

Primary Fisher BRK cells were prepared from 6 day old baby rats and cultured in Dulbecco's modified eagle medium (DMEM) with 10% fetal bovine serum as described previously (White et al. 1991). Plasmids were
20 transfected by electroporation, and cells were maintained in DMEM with 10% fetal bovine serum. The transformed BRK cell line, p53A, was derived from transfection of pCMVE1A and pLTRcGval135 plasmids (Debbas and White 1993). The 19K1 and 4B cell lines
25 were derived from transfecting pCMV19K and pSVBcl-2, respectively, and a neomycin-resistance plasmid, pSVneo, by electroporation (Debbas and White 1993) (Chiou et al., 1994; Sabbatini et al., 1995). Control cell line, p53An1, was derived from p53A cells containing only the neomycin-resistance plasmid (Debbas and White 1993).
30 The LTR.1A cells were generated by transfecting primary BRK cells with 10 µg of LTR H-myc, which expresses human c-Myc, and pLTRcGval135 as described previously

(Sakamuro et al. 1995).

Antibodies

5 SMP14, a mouse monoclonal anti-Mdm2 antibody (Santa Cruz Biotechnology, Inc, Santa Cruz, CA) was used at 1:1000 (Western) or 1:100 (immunofluorescence) to detect Mdm2. Clone 248, a murine-specific anti-p53 mouse monoclonal antibody which was kindly provided by Dr. A. Levine was used at 1:5000 to detect p53. Mouse
10 monoclonal anti-p21^{WAF1} (Ab-4, Calbiochem, San Diego, CA) was used at 1:50 dilution to detect p21^{WAF1}. A rabbit polyclonal anti-Bax (P-19, Santa Cruz) was used at 1:100 to detect Bax and a mouse monoclonal anti-actin antibody (Amersham) was used at 1:1000 dilution to detect actin.
15 Anti-HA mouse monoclonal antibody (Babco, Richmond, CA) at 1:1000 dilution was used to detect ectopically expressed p300. A mouse monoclonal anti-E1A antibody M73 (Calbiochem) was used at 1:1000 dilution to detect E1A. A polyclonal anti-E1B 19K antibody was used at
20 1:10,000 to detect transiently expressed E1B 19K.

Cell viability and morphology

Cell viability was measured by trypan blue staining. Cells were trypsinized, centrifuged,
25 resuspended in PBS, and counted using a hemocytometer after diluting in trypan blue. Apoptotic morphology was assessed by scoring for cells that are positive by fluorescence microscopy and are rounded and non adherent. Cell morphology was also documented by
30 photography at 100 x magnification on a Nikon phase-contrast microscope and camera using Kodak film.

Western Blot Analysis

Cells were incubated at 37.5°C and 32°C for various time intervals prior to lysing in buffer containing 4% SDS and 5% β -mercaptoethanol. Equal amounts of protein (20-30 μ g) were electrophoresed on 7.5-12% SDS-PAGE gels and transferred to PVDF membranes. Membranes were blocked for 15 minutes at room temperature in 5% nonfat Carnation dry milk in PBS containing 0.1% Tween-20 (PBST). Membranes were incubated with primary antibody followed by washes in PBST and then incubated with horseradish peroxidase (HRP)-conjugated sheep anti-mouse IgG or donkey anti-rabbit IgG monoclonal antibody at 1:2000 dilution. After several washes, blots were developed using the ECL chemiluminescence detection kit according to manufacturer's recommendations (Amersham LIFE SCIENCES, Arlington Heights, IL).

Indirect immunofluorescence and microscopy

Cells were fixed in cold methanol prior to incubation with primary antibodies. Subsequent to several washes in PBS, cells were incubated with rhodamine-conjugated goat anti-mouse antibody or fluorescein-conjugated goat anti-rabbit antibody. Cells were then washed, mounted and analyzed by fluorescence microscopy using a Nikon FXA epifluorescence microscope.

Northern Blot Analysis

Cytoplasmic RNA was extracted from p53A and 19K1 cells using the NP-40 lysis protocol as previously described (Muraoka et al. 1996). Northern blots were performed by loading 30 μ g of cytoplasmic RNA on a formaldehyde gel and blotted as previously described (Muraoka et al. 1996) using Hybond-N membranes

(Amersham) for transfer. The membrane was hybridized with a random-primed labeled murine *mdm2*-specific probe in ExpressHyb Hybridization solution (Clontech, Palo Alto, CA) at 65°C. Blots were washed in 0.1X SSC and 0.1% SDS at 50°C and exposed to film for 2 days.

Transient Transfections

The indicated amount of plasmid DNA was transfected by electroporation. After transfection, cells were incubated at 37.5°C for 24 hours to recover, and were then shifted to 32°C for 24 hours before analysis for viability by trypan blue staining and apoptotic morphology. For p300 transfections, p53A cells were electroporated with 12 or 24 µg of pCMVβ alone, pCMVβp300 or pCMVp300Δ30 mutant (deletion in E1A-interacting domain) (Arany et al. 1994). For transient expression of E1A proteins, LTR.1A cells were electroporated with 10 µg of pCMVβ alone, pCMV12SE1A, pCMV12S.RG2, or pCMV12S.YH47.928 (Wang et al. 1992). For Mdm2 transient expression, p53A cells were electroporated with 5-10 µg of pCMVβ alone, pCMVβ-X₂ (wild-type Mdm2), and pCMVβ-ΔXM (mutant Mdm2 with deletion of the N-terminal p53-binding domain) (Haupt et al. 1997).

Stable transfections

Cells were transfected by electroporation using 2 µg of pCMVβ alone or pCMVβ-X₂ and 200 ng of pSVneo along with 10 µg of salmon sperm DNA. Cells were selected in complete media containing 0.5 mg/ml G418 (Sigma) for a week and then incubated at the permissive temperature to select for cells stably expressing Mdm2. Two independent pools of p53A cells stably expressing wild-

type Mdm2 were analyzed.

Yeast two-hybrid system for identifying E1B 19K binding proteins

5 Procedures for utilizing the two-hybrid system to
identify E1B 19K binding proteins were described
previously (Han et al., 1996). A HeLa cDNA library was
constructed in the pGAD-GH vector and screened against
pGBT9-E1B 19K. Plasmids were transformed into *S.*
10 *cerevisiae* YGH1 cells and positive clones were selected
based on growth in the absence of histidine and
production of β -galactosidase. False-positive clones
were eliminated by testing for interactions with an
irrelevant hydrophobic protein (Apc-2) and the empty
15 pGBT9 vector. Missense and deletion mutants of E1B 19K
tested for interaction with BP-1 were described
previously (Han et al., 1996, *supra*). pGBT8-Bcl-2 (20)
and pGBT9-Bax (26) were also described previously.
pGBT8-Bcl- χ_L was kindly provided by Dr. G. Nuñez (Univ.
20 of Michigan, Ann Arbor, MI).

The full length transcript, *btf_s*, was identified by
screening a λ -cDNA library prepared from HeLa cells
(Stratagene, La Jolla, CA) using conventional
techniques. The cDNA sequence of *bp-1* and the
25 subsequent full length *btf* were analyzed using Sequenase
2.0 (US Biochemical, Cleveland, OH) according to the
manufacturer's specifications and later confirmed by
fluorescent terminator cycle sequencing using an
automated 377 DNA sequencer (Perkin-Elmer, Applied
30 Biosystems, Foster City, CA).

Plasmid construction

A PCR product of *btf_s* from the λ -screen was digested

with *XmaI* and *NotI* (blunt) and ligated into both pGAD-GH cut with *XmaI*-*XhoI* (blunt) and pGBT9 cut with *XmaI*-*SalI* (blunt). To prepare pcDNA3-Myc-Btf₈ for mammalian expression and *in vitro* translation, oligonucleotides encoding a Myc epitope with flanking *KpnI* and *XmaI* sites were annealed with a PCR cloned fragment of *btf₈* from bluescript digested with *XmaI* and *NotI*, and pcDNA3 (Invitrogen, San Diego, CA) digested with *KpnI* and *NotI*. The resulting plasmid encodes a Myc epitope at the N-terminal end of Btf₈. Btf₈ and two deletion mutants were fused in frame with the GAL-4 DNA-binding domain in the pml vector for use in transcriptional reporter assays (kindly provided by Dr. C. Abate-Shen, Center for Advanced Biotechnology and Medicine, Piscataway, NJ).

The full-length *btf₈* construct was prepared by digesting pGBT9-Btf₈ with *XmaI* and *PstI* and ligating into the same sites in the pml vector. The N552 and C210 mutants were generated by digesting pGBT9-Btf₈ with *EcoRI*-*PstI* and *XmaI*-*BamHI*, respectively, and ligating into the same restriction sites within pml. *btf₈* was also cloned into pIRES-EGFP (Clontech, San Francisco, CA) for cell cycle analysis of Btf₈ expressing cells. The construct was prepared by digesting pcDNA3-Myc-Btf₈ with *SmaI* and *NotI* and ligating to pIRES-EGFP cut with *EcoRV* and *NotI*. The pcDNA3-Bcl-2 plasmid was prepared by ligating an *EcoRI*-*XhoI* fragment from pSFFV-Bcl-2 (Hockenberry et al., 1990) (provided by Dr. S. Korsmeyer, Washington University School of Medicine, Saint Louis, MO) into pcDNA3. The C-terminally V5/His tagged E1B 19K was prepared by TA cloning into pcDNA3.1/V5/His-TOPO (Invitrogen, San Diego, CA) according to manufacturer's specifications.

Cell lines

The HeLa cells were maintained in culture in Dulbecco modified Eagle medium (D-MEM) with 10% fetal bovine serum (FBS) at 37°C / 5% CO₂. Stable HeLa cell lines containing Bcl-x_L were prepared by electroporating 1 µg of pcDNA3-Flag-Bcl-x_L (Merino et al., 1995), provided by Dr. G. Nuñez (Univ. of Michigan, Ann Arbor, MI), into HeLa cells and selecting with 1.2 mg/ml geneticin. Expression of Bcl-x_L was verified by immunofluorescence.

Northern blot for btf expression

Northern blot analyses were performed using commercially available blots of multiple tissues or cancer lines (Clontech, San Francisco, CA). Each lane contained 2 µg of the indicated poly A⁺ RNA. The btf_s probe was prepared by random priming using a purified XmaI-XhoI fragment from pGAD-GH-Btf_s. Human β-actin cDNA supplied with the blots were used as a control probe to confirm equal mRNA loading. Hybridization was performed using ExpressHyb solution (Clontech, San Francisco, CA) according to manufacturer's specifications. Bands were visualized by autoradiography and using a PhosphorImager (Molecular Dynamics, Sunnyvale, CA).

In vitro binding assay

Binding reactions were performed by combining ³⁵[S]-methionine-labeled in vitro translated Myc-Btf_s or Myc-Bax (Han et al., 1996) with V5/His-E1B 19K, Bcl-2, Flag-Bcl-x_L, or luciferase (Promega Corp., Madison, WI) prepared using the TNT T7 reticulocyte lysate system (Progema Corp., Madison, WI) according to manufacturer's specifications. Samples were incubated with anti-Myc

monoclonal antibody (Oncogene Research Products, Cambridge, MA) in 500 μ l of NETN buffer [20 mM Tris (pH 8.0), 100 mM NaCl, 1 mM EDTA, and 0.2% NP-40] for 1.5 hrs at 4°C followed by protein A-sepharose. All samples were then washed three times in NETN buffer, resuspended in 2X Laemmli buffer, boiled for 5 minutes, and analyzed by SDS-PAGE. Gels were fixed in 50% methanol, 10% acetic acid for 1 hour, dried and then visualized by autoradiography.

DNA binding assay

³⁵[S]-Methionine labeled *in vitro* translated Myc-Btf_s and E1B 19K were prepared from 1 μ g of pcDNA3-Myc-Btf_s and pcDNA3-E1B 19K (Han et al., 1996, supra) using the Twt T7 reticulocyte lysate system. The proteins were incubated with 200 μ l of native DNA-cellulose (Pharmacia Biotech, Piscataway, NJ) in 500 μ l of NETN buffer for 2 hrs at 4°C. The relative levels of Btf_s and E1B 19K production were determined by immunoprecipitation with anti-Myc (Oncogene Research Products, Cambridge, MA) or anti-E1B 19K (White and Cipriani, 1989) in NETN buffer followed by protein A-sepharose and then washed in NETN. Samples were resolved by SDS-PAGE and examined by autoradiography as described above.

Indirect immunofluorescence

HeLa cells were electroporated with 15 μ g of pcDNA3-Myc-Btf_s along with 15 μ g of either pCMV-E1B 19K (White and Cipriani, 1989, supra) or pcDNA3-Bcl-2. Total amount of DNA was kept constant using empty pcDNA3 vector. Cells grown on glass coverslips were stained 24 hours post-transfection as described previously (Perez

and White, 1998). Briefly, the cells were fixed with 2% paraformaldehyde in Phosphate Buffered Saline (PBS), and then permeabilized with 0.5% Triton X-100 in PBS. The cells were double labeled with an anti-Myc mouse monoclonal antibody (Oncogene Research Products, Cambridge, MA) along with either an anti-E1B 19K rabbit polyclonal (White and Cipriani, 1989, supra) or an anti-Bcl-2 hamster monoclonal antibody (PharMingen, San Diego, CA). Antibody complexes were visualized with rhodamine-conjugated goat anti-mouse antibody along with the fluorescein-conjugated antibodies goat anti-rabbit or goat anti-hamster (Jackson ImmunoResearch Laboratories, Inc., West Grove, PA). Immunocytochemistry was also performed in HeLa-Bcl-x₂ cells electroporated with 15 µg of pcDNA3-Myc-Btf_s. These cells were stained for the Myc epitope as described above. Expression of Flag-tagged Bcl-x_L was confirmed by staining separate coverslips with anti-Flag M5 monoclonal antibody (Scientific Imaging Systems, New Haven, CT) and rhodamine-conjugated goat anti-mouse. The cells were visualized by epifluorescence with a Nikon FXA microscope (Nikon Inc., Garden City, NY).

Transcription assays

Transcriptional reporter assays were performed as described previously (Catron, et al., 1995). HeLa cells were plated onto 60 mm tissue culture dishes and grown to 50-75% confluency. The cells were then transfected with 2.5 µg of a GAL-4 luciferase reporter construct (kindly provided by Dr. C. Abate-Shen, Center for Advanced Biotechnology and Medicine, Piscataway, NJ) along with 2.5 µg of both a pml vector (Btf_s, Btf_s-N552 or Btf_s-C210) and a bcl-2 family gene (pCMV-E1B 19K,

pcDNA3-Bcl-2, or pcDNA3-Flag-Bcl-x_L). DNA concentrations were kept constant using appropriate empty vectors, either pml or pcDNA3. The cells were transfected using SuperFect (Qiagen, Valencia, CA) according to the manufacturer's specifications and harvested 24 hours post-transfection. Expression of mutant and wild-type Btf₃ proteins were verified by immunofluorescence using an antibody against the GAL-4 DNA-binding domain (Clontech, San Francisco, CA). Luciferase activity was determined using the Luciferase Assay System (Promega Corp., Madison, WI) measured in a scintillation counter, and then normalized for protein concentrations measured by a Bradford assay. Values were graphed as a percent of the negative control (empty pml, empty pcDNA3, and GAL-4 luciferase).

Transformation assay

Transformation assays of baby rat kidney (BRK) cells were performed as described previously (White et al., 1992). Briefly, BRK cells prepared from 6-day-old Fisher rats were electroporated with carrier DNA along with linerized test DNA (15 µg pCMV-E1A (White et al., 1991), 15 µg pCMV-E1B 19K or pCMV-p53DD (Shaulian, et al., 1992), and 45 µg pcDNA3-Myc-Btf₃). DNA concentrations were kept constant using appropriate empty vectors. The cells were cultured for 3 to 4 weeks at 38.5°C in D-MEM supplemented with 5% FBS, and then stained with Giemsa. Foci were counted from four dishes per condition.

Cell cycle analysis

HeLa cells were electroporated with 20 µg pIRES-EGFP- Btf₃ or empty pIRES-EGFP vector in combinations

with 10 μ g pCMV-E1B 19K or empty pcDNA3 vector. Cells were harvested 48 and 72 hours post-transfection and fixed in 2% paraformaldehyde in PBS for 30 min at 4°C. The cells were washed and then stained with PBS containing 1 μ g/ml propidium iodide, 250 μ g/ml RNase A, and 0.1% Tween 20 for at least 30 min at room temperature. The fluorescent intensities for EGFP and propidium iodide were analyzed by FACS (EPICS PROFILE-II; Coulter, Miami, FL). In addition, the live transfected cells were stained with Hoechst dye to visualize the DNA and photographed with a Nikon FXA microscope (Nikon Inc., Garden City, NY).

The following examples are provided to illustrate particular embodiments of the invention. They are not intended to limit the invention in any way.

EXAMPLE I

Mdm2 expression regulates p53-dependent apoptosis in BRK cells

BRK cells were transformed by a temperature sensitive p53 (val135) mutant and either adenovirus E1A (p53A) or c-Myc (LTR.1A). As previously reported, both types of cell lines expressed the temperature sensitive p53 (val135) mutant in which p53 is in the wild-type conformation at the permissive temperature of 32°C and in the mutant conformation at the nonpermissive temperature of 37.5°C (Debbas and White 1993; Sakamuro et al. 1995). There are two striking differences between E1A + tsp53(val135) transformed (p53A) and myc + tsp53(val135) transformed (LTR.1A) cell lines. First, BRK cells expressing E1A undergo massive apoptosis at

the permissive temperature (Chiou et al. 1994; Debbas and White 1993; Sabbatini et al. 1995) whereas the Myc expressing cells undergo a wave of apoptosis within the first 24 hours at the permissive temperature but are predominantly resistant to apoptosis (Sakamuro et al. 1995). Second, E1A expressing cells maintain high levels of p53, whereas the p53 levels are down-regulated at 32°C in Myc expressing cells (Debbas and White 1993; Sakamuro et al. 1995). Thus, high levels of p53 correlate with augmentation of the apoptotic response in E1A expressing p53A cells. Since E1A stabilizes p53 (Chiou et al. 1994; Lowe and Ruley 1993) and this activity requires that E1A binds to p300 (Chiou and White 1997), this suggests a possible function for sequestration of p300 and induction of p53 levels. Furthermore, recent reports demonstrate the requirement for p300 in p53 transactivation (Gu et al. 1997; Lill et al. 1997; Scolnick et al. 1997), and a role for Mdm2 in promoting p53 degradation (Haupt et al. 1997; Kubbutat et al. 1997). The possible role of p300 in the Mdm2-dependent negative-feedback loop and the control of p53-dependent apoptosis was further investigated.

In accordance with the present invention it was determined whether the levels of p53-inducible gene products Mdm2, p21^{WAF1} and Bax were affected by the p300-binding protein E1A. Western blot analysis shows that p53 levels were up-regulated in E1A transformed cells and down-regulated in Myc transformed cells at 32°C (Fig. 1A) as previously reported (Debbas and White 1993; Sakamuro et al. 1995). A lower band which may represent a proteolytic cleavage product of p53 was also observed in p53A cells as apoptosis progressed at the permissive temperature. See Figure 1A. High p53 levels correlated

with apoptosis in p53A cells. See Figure 3A and Figure 4, top panel. Endogenous Mdm2 protein was initially present but was down-regulated at 32°C in p53A cells as shown by Western blot analysis (Figure 1) and immunofluorescence (Figure 2, upper panel). Western blot analysis of Mdm2 in p53A cells at the permissive temperature showed a lower band around 50 kDa recognized specifically by the anti-Mdm2 monoclonal antibody (data not shown). This lower band may represent a cleavage product of Mdm2. However, this band disappeared as cells were incubated at the permissive temperature. Mdm2 down-regulation was surprising since *mdm2* is a p53-regulated gene and suggested that p300 may be necessary for Mdm2 transactivation by p53.

In c-Myc transformed LTR.1A cells, Mdm2 was up-regulated and p53 levels were low at 32°C. See Figure 1B and Figure 2, lower panel). Therefore Myc, but not E1A expressing BRK cells demonstrate p53-dependent induction of *mdm2*. Furthermore, high levels of Mdm2 in Myc transformed BRK cells may have promoted p53 degradation as shown in Figure 1A. The LTR.1A cells initially undergo apoptosis but rapidly become resistant to p53-mediated cell death at 32°C (Figure 3, bottom panel and Figure 4, bottom panel) which correlated with the down-regulation of p53 and up-regulation of Mdm2.

Mdm2 is only one of many p53-inducible genes that collectively act to modulate the physiological response to p53 induction. Therefore, p21^{WAF1} and Bax levels were also examined for transactivation in p53A and LTR.1A cells. In p53A cells, Bax and p21^{WAF1} levels were induced at 32°C even in the presence of E1A and inhibition of p300 as shown in Figure 1A and Figure 1B (Han et al. 1996; Sabbatini et al. 1995). LTR.1A cells down-

regulated Bax but up-regulated low levels of p21^{WAF1} at 32°C. See Figure 1. Similar results (data not shown) were obtained with independent E1A + tsp53(val135) and c-Myc + tsp53(val135) transformed clones (Debbas and White 1993; Sakamuro et al. 1995). These results indicate that E1A and Myc differentially alter p53-dependent gene expression.

In order to determine whether the absence of Mdm2 protein in p53A cells reflected a transcriptional event, Northern blot analysis was performed using cytoplasmic RNA extracted from E1A transformed p53A cells incubated at the permissive temperature for the indicated time intervals. See Figure 5. Two transcripts of 3.3 and 1.7 kb hybridized to a mdm2-specific probe. Both transcripts, particularly the 3.3 kb, were down-regulated to some extent as cells were incubated at 32°C. These results suggest that p300 may be required for Mdm2 transactivation, but may be dispensable for Bax and p21^{WAF1} regulation.

p300 is required for the up-regulation of Mdm2 by p53 and inhibition of p53-mediated apoptosis

We next attempted to rescue p53A cells from apoptosis at the permissive temperature by ectopically expressing p300. Vector alone, wild-type p300 and a functional but truncated form of p300 with a deletion of E1A binding domain (Arany et al. 1994), were transiently transfected in p53A cells. p53-dependent apoptosis was examined at the permissive temperature 48 hours post-transfection. p53A cells transiently expressed the exogenous wild-type p300 and the E1A binding mutant form of p300, neither of which were detected in mock transfected cells. See Figures 6 and 7. The up-

regulation of Mdm2 in p300 transfected cells was not detected by Western blot analysis above endogenous levels (Fig. 6) because the transfection efficiency was low in transient assays. However, the up-regulation of Mdm2 in both wild-type and E1A-binding mutant of p300 was easily observed in 90% of transfected cells by indirect immunofluorescence when individual transfected cells were examined as shown in Figure 7, lower panel, arrows. Furthermore, ectopic expression of wild-type and mutant p300 rescued cells from apoptosis as cell viability was elevated as much as 20-40% above mock transfected cells. See Figure 8. The mutant form of p300 which was unable to bind E1A, protected cells better than wild-type p300, presumably because it evades E1A sequestration.

In addition, we determined the percentage of cells with apoptotic morphology that were ectopically expressing wild-type p300 ectopically or the E1A-binding mutant of p300 following transient transfection with 24 μ g of plasmid DNA. Apoptotic morphology was determined by counting cells that were rounded in shape and detaching from the dish. The percentage of cells with apoptotic morphology transiently expressing a control protein, a fragment of the lamin A protein (Rao et al. 1996), compared to p300 and mutant p300 transfected p53A cells at the permissive temperature was determined by indirect immunofluorescence. See Figure 9. Only 4-8% of the cells expressing wild-type p300 and mutant p300, respectively, have apoptotic morphology compared to 62% cells expressing the lamin fragment (Fig. 9). These results reaffirm that expression of wild-type p300 or the E1A-binding p300 mutant is sufficient to rescue cells from p53-mediated apoptosis.

In order to determine whether LTR.1A cells can be made susceptible to p53-mediated death, wild-type E1A (12S E1A) was transfected into these cells. Transient expression of 12S E1A in LTR.1A cells, see Figure 10, promoted apoptosis at the permissive temperature as shown in Figure 11. In addition, Mdm2 levels were down-regulated in cells transfected with 12S E1A as detected by Western blot analysis (Fig. 10). Since E1A can also bind to Rb, we transiently transfected LTR.1A cells with 12S E1A mutant (12S.RG2) that is unable to bind to p300 but retains its ability to bind to Rb, and a 12S E1A mutant (12S.YH47.928) that binds to p300 but not to Rb (Wang et al. 1992), and examined p53-dependent apoptosis at the permissive temperature. Cells transfected with 12S.RG2 did not down-regulate Mdm2 (Fig. 10) and were resistant to p53-dependent apoptosis (Fig. 11). In contrast, cells transfected with 12S.YH47.928 down-regulated Mdm2 (Fig. 10) and were susceptible to p53-dependent apoptosis at the permissive temperature (Fig. 11). These results suggest that the ability of E1A to bind to p300 and inhibit Mdm2 induction correlates with E1A-mediated, p53-dependent apoptosis.

Overexpression of Mdm2 inhibits p53-mediated apoptosis

To determine whether Mdm2 expression can bypass the requirement for p300 cotransactivation and rescue p53A cells from p53-mediated death, p53A cells were transiently transfected with expression plasmids encoding wild-type Mdm2 and a truncated form of Mdm2 lacking the N-terminal p53-binding domain (Haupt et al. 1997). Transient overexpression of wild-type and mutant Mdm2 were detected by Western blot analysis in p53A cells (Fig. 12). Transient transfection of cells with 5

or 10 μ g wild-type Mdm2 rescued cells from p53-dependent apoptosis at the permissive temperature (Fig. 13). The truncated form of Mdm2 rescued cells from apoptosis less efficiently than the wild-type protein (Fig. 13), presumably because it was not able to bind to p53 and promote its degradation. Furthermore, we examined the cell morphology of p53A cells expressing a wild-type and mutant Mdm2 plasmid DNA as compared to the morphology of cells expressing a control lamin fragment incubated at the permissive temperature using indirect immunofluorescence (Fig. 14). The percentage of apoptotic cells expressing wild-type and mutant Mdm2 was only 10% and 30%, respectively, as compared to 56% apoptosis in cells expressing the control lamin fragment (Fig. 14). The Mdm2 mutant, however, possessed greater apoptotic suppressing activity than the control lamin fragment which may indicate that Mdm2 can suppress apoptosis by a mechanism independent of p53 binding. These results demonstrate that overexpression of Mdm2 can rescue E1A-expressing cells from p53-mediated apoptosis by either promoting p53 degradation or by inhibiting its activity by direct interaction.

E1B 19K or Bcl-2 can bypass E1A inhibition of p300 and restore mdm2 transactivation.

Another mechanism for suppressing p53-dependent apoptosis is through expression of Bcl-2 or E1B 19K, which function in part by binding to Bax and inactivating its pro-apoptotic function. To test if Bcl-2 or E1B 19K could also function to suppress apoptosis by influencing p53 target gene expression, we examined p53-inducible gene products, Mdm2, p21^{WAF1} and Bax, in p53A cells stably expressing adenovirus E1B 19K (19K1)

or its human homolog Bcl-2 (4B).

As reported previously, the expression of 19K or Bcl-2 abrogates p53 dependent apoptosis at 32°C (Chiou et al., 1994a; Sabbatini et al., 1995a; Han et al., 1996). Western blot analysis indicated that p21^{WAF1} and Bax levels increased at 32°C regardless of whether or not 19K or Bcl-2 were expressed and the levels of p53 remained high. See Figures 15A, 15B and 15C. However, the presence of 19K or Bcl-2 dramatically up-regulated Mdm2 levels, whereas the control E1A + tsp53 (val135-transformed p53A cells transfected with vector alone, (p53An1) down regulated Mdm2 as expected (Fig. 15). This suggested that E1B 19K or Bcl-2 could restore p53-dependent mdm2 expression in p53A cells.

To determine whether the expression of E1B 19K or Bcl-2 was sufficient for up-regulation of Mdm2 levels, we transiently transfected 19K or Bcl-2 in p53An1 cells and examined Mdm2 levels at 32°C. Western blot analysis (Fig. 16), and indirect immunofluorescence indicated that Mdm2 levels were up-regulated when either 19K or Bcl-2 was expressed transiently.

To determine whether expression of E1B 19K upregulates Mdm2 at the transcriptional level, we performed Northern blot analysis using cytoplasmic RNA from cells expressing E1B 19K (19K1) incubated at 32°C for the indicated time intervals (Fig. 17). As shown in Figure 17, mdm2 mRNA expression was up-regulated after 48 hour incubation at the permissive temperature. This suggested that E1B 19K and Bcl-2 may restore mdm2 transactivation by p53, which would otherwise be inhibited by E1A binding to p300. In addition, previous studies have shown that Bax, and p21^{WAF1} mRNA and protein were also up-regulated in 19K1 cells incubated at the

permissive temperature. Therefore, the presence of 19K did not affect the expression of Bax or p21^{WAF1}, however, E1B 19K can directly bind to Bax and inhibit its function. E1B 19K and Bcl-2 have been shown to relieve transcriptional repression by E1A and p53, which may be responsible for inhibition of apoptosis and Mdm2 may be a physiological target of this activity.

Discussion

E1A + tsp53(vall135) transformed p53A cells were unable to transactivate *mdm2* as a result of E1A inhibition of p300, indicating that p53 requires p300 to transactivate *mdm2*. See Figure 18. Furthermore, low Mdm2 levels in p53A cells relieve the negative-feedback regulation of p53 resulting in high p53 accumulation leading to apoptosis. In contrast, c-Myc + tsp53(vall135) transformed LTR.1A cells transactivate Mdm2 in the absence of E1A inhibition of p300 (Fig. 18). Up-regulation of *mdm2* inhibits p53 function and promotes p53 degradation (Haupt et al. 1997; Kubbutat et al. 1997), which in turn inhibits p53-mediated cell death resulting in growth arrest in LTR.1A cells. Cellular levels of p53 control the response of the cells such that lower levels of p53 induce growth arrest and higher levels induce apoptosis (Chen et al. 1996). Therefore, it may be concluded that p300 regulation of Mdm2 levels determines whether the physiological response to p53 is growth arrest or apoptosis.

Interestingly, p300 cotransactivation is not required for the other p53-inducible genes Bax and p21^{WAF2}. It has previously been reported that E1A inhibits p53-mediated transactivation using promoter reporter assays (Steegenga et al. 1996). Our studies,

however, demonstrate that E1A specifically inhibits endogenous *mdm2* transactivation and not other p53-inducible genes such as *bax* and *p21^{WAF1}*. p300/CBP functions by interacting not only with p53, but also with other factors such as the TAFs (Thut et al. 1995), TBP (Abraham et al. 1993), CREB (Chrivia et al. 1993; Kwok et al. 1994), c-Jun/v-Jun (Bannister and Kouzarides 1995), c-Myb/v-Myb (Dai et al. 1996), c-Fos (Bannister and Kouzarides 1995) and others. p53-binding alone may not regulate the specificity of p300 cotransactivation, however, as numerous other cellular interactions may determine the specificity transactivation, which may explain differential regulation of p53-inducible genes by p300.

p300/CBP transcriptional coactivators have been shown to have histone acetyltransferase activity which can modify chromatin structure and enhance gene expression (Bannister and Kouzarides 1996; Ogryzko et al. 1996). Recently, p300 has been shown to acetylate p53 itself (Gu and Roeder 1997). The acetylation of p53 increases the binding activity of p53 to specific consensus sequences. In vivo, this activity may be required for facilitating p53 tetramer interactions with the DNA template, thereby promoting *mdm2* transactivation.

Mutations that inactivate p300 have been described in colorectal and gastric carcinomas (Muraoka et al. 1996), which suggests that p300 functions as a negative regulator of cell growth. Missense mutations of p300 coupled with the deletion of the second allele of the gene were observed in these carcinomas. These observations suggest that inactivation of p300 may play a role in the development of cancer. E1A inhibition of

p300 transactivation may be one mechanism whereby this viral protein promotes cellular transformation. Moreover, p300 inactivation may render cells susceptible to agents that induce p53-mediated cell death such as UV and ionizing radiation. It is intriguing to speculate that acetyltransferase inhibitors specific for p300 may be used in cells that do not respond to chemotherapy regardless of having wild-type p53.

EXAMPLE II

As discussed previously, apoptosis is a genetically controlled process of cell suicide that plays a critical role in maintaining homeostasis and preventing disease. Disruption of apoptosis leads to impaired development, cancer, neurodegenerative and autoimmune diseases, and sustained viral infection. The regulation of apoptosis is a precarious balance between factors that promote survival and those responsible for initiating and executing cell death. One major advance toward the understanding of apoptosis regulation has been the characterization of the Bcl-2 family (White, E., 1996). This family consists of highly conserved proteins with opposing biological function. Anti-apoptotic Bcl-2 family members, such as Bcl-2 and Bcl-x_L, inhibit apoptosis triggered by many circumstances, including TNF- α , Fas, UV radiation, chemotherapeutic drugs, and growth factor/hormone withdrawal. In contrast, pro-apoptotic Bcl-2 family members, such as Bax, Bak, and Nbk/Bik, induce cell death in numerous model systems. The adenovirus E1B 19K protein cooperates with E1A in transformation assays and is a viral homologue of mammalian Bcl-2. Expression of E1B 19K, or Bcl-2, inhibits E1A-induced, p53-mediated apoptosis (Chiou et

al, 1994; Han et al., 1996). Like Bcl-2, E1B 19K interacts with and antagonizes several pro-apoptotic family members, including Bax (Han et al., 1996), Nbk/Bik (Boyd, et al., 1995), and Bak. However, it is still unclear how these proteins affect cell survival or death and whether binding to proteins unrelated to the Bcl-2 family contribute to apoptosis or other cellular processes.

While there has been some debate concerning the biochemical function of E1B 19K and other Bcl-2 family members in the regulation of apoptosis, recent studies have revealed several possible mechanisms. Clearly, interactions between Bcl-2 family members play an integral part in the regulation of apoptosis and transformation. A common feature of the Bcl-2 family is the occurrence of protein-protein interactions between the anti- and pro-apoptotic proteins, the ratio of which controls the fate of the cell (Boise et al., 1993; Oltavai et al., 1993). Structural and biochemical studies of Bcl-2 family members point to a possibility that these proteins form ion channels (Antonsson et al., 1997). In addition, Bcl-2 family members bind to several unrelated proteins including R-ras, Nip-1, -2, -3, Ced-4-like proteins, Bag-1, Lamin A/C, and p28Bap31. Although the functional significance of some of these interactions are not yet known, these findings suggest that Bcl-2 regulates multiple signaling pathways that influence apoptosis.

There is growing evidence from several independent studies that Bcl-2-related proteins can trigger changes in gene expression which may or may not be related to their role in apoptosis regulation (Linette et al., 1996; Miyashita et al., 1997). Initial studies of E1B

19K mutant viruses raised the possibility that 19K functions to dampen transcriptional activation by E1A. Subsequently, it has been shown that E1B 19K and Bcl-2 alleviate the *trans*-repressive activity of E1A and p53. This derepression of transcription has been correlated with an up-regulation of one transcriptional target of p53, Mdm-2 as described in Example I. Since Mdm-2 inhibits p53 transcriptional activity and apoptosis, this model provides an alternative mechanism by which E1B 19K and Bcl-2 can regulate apoptosis. Furthermore, Bcl-2 family members may actually control several additional transcription factors including NF- κ B, NFAT (nuclear factor of activated T cells), c-Jun, and the glucocorticoid receptor, thus indicating that Bcl-2 and its related proteins can have multiple effects on gene expression that may contribute to apoptosis.

Compared to apoptosis regulation, relatively little is known concerning the gene regulatory function of the Bcl-2 family. In accordance with the present invention, a novel transcriptional repressor, Btf, which contributes to the modulation of transcription by Bcl-2 related proteins is provided. Btf was identified in a yeast-two hybrid screen against E1B 19K and subsequently shown to also interact with Bcl-2 and Bcl-x_L. These Bcl-2 family members inhibit the translocation of Btf to the nucleus and abrogate its transcriptional activity. The data presented herein reveal that sustained over-expression of Btf induces apoptosis and suppresses transformation by E1A and E1B 19K or mutant p53. Thus, the interaction Btf provides yet another novel pathway by which the Bcl-2 family can regulate transcription and control apoptosis.

Yeast Two Hybrid Assay

In order to identify novel cellular proteins that interact with E1B 19K, we screened a HeLa cDNA library using the yeast two-hybrid assay. The library was constructed in the pGAD-GH plasmid in which the cDNA sequences were fused to the GAL-4 activation domain, and the bait gene, E1B 19K, was fused to the GAL-4 DNA-binding domain in the pGBT9 vector (Han et al., 1996). The plasmids were transformed into the YGH1 yeast strain and screened for GAL-4 inducible phenotypes, namely growth in the absence of histidine and production of β -galactosidase. Three million transformants were screened yielding seven clones (BP-1 to BP-7) that specifically interacted with 19K and not with the pGBT9 vector alone or with an irrelevant protein, Apc-2. BP-2, -3, and -4 were previously reported as Lamin A/C, Bax, and Nbk/Bik, respectively. Here, we describe one of the novel E1B 19K associated proteins, BP-1. A 1.5 kB cDNA containing *bp-1* was isolated seven times during the two-hybrid screen. Since Bcl-2 family members are highly homologous and frequently interact with the same cellular proteins, we tested BP-1 for interaction with Bcl-2-related apoptosis regulators. In addition to binding E1B 19K, BP-1 interacted with other related proteins, Bcl-2 and Bcl-x_L, but not to the pro-apoptotic family member, Bax (Fig. 19A).

The interaction between anti- and pro-apoptotic Bcl-2 family members generally occurs via their conserved domains, designated Bcl-2 homologous regions 1, 2, 3, and 4 (BH1, BH2, BH3, and BH4). BH1-3 are in close proximity forming a hydrophobic cleft that is required for dimerization. To determine the regions of E1B 19K that are required for interaction with BP-1, we

tested its ability to bind a series of E1B 19K missense and deletion mutants (See Figures 19 A,B & C). These mutants have been characterized previously for binding to other E1B 19K binding proteins, including Bax, Nbk/Bik, Lamin A/C, and Ced-4. Thus, these experiments allow us to compare the binding requirements within the E1B 19K protein against other 19K-associated proteins.

The E1B 19K protein may be divided into three regions: a moderately conserved N-terminus which includes BH3, a highly conserved central region containing BH1, and a poorly conserved C-terminus. It is important to note that most of the deletion mutants tested Δ N30, Δ N64, Δ N87, Δ C93, Δ C70, 30-146, 30-93, and 64-136) failed to interact with BP-1 or with Bax, Nbk/Bik, and Lamin A/C, and that this may be due to abnormal protein folding or masking of the binding site(s) (Fig. 19B). Like Bax, Nbk/Bik, and Lamin A/C, binding of BP-1 was retained in C146 which contains both BH1 and BH3. The small E1B 19K fragment 19-57 containing only BH3 was also able to bind BP-1 (Fig. 19B). This fragment is also sufficient for binding to Bax as well as Ced-4. Surprisingly, we found that another small region of E1B 19K, C36, also retained binding to BP-1 (Fig. 19B). This region did not bind to other E1B 19K-associated proteins, Bax, Nbk/Bik, Lamin A/C, or Ced-4. The binding of BP-1 with the E1B 19K mutants C36 and 19-57, suggests that the region of 19K immediately adjacent to BH3 (19-36) may be sufficient for the interaction. This would represent a unique domain that contributes to protein-protein interactions and may correspond to the E1B 19K BH4, although homology is weak.

We further addressed the binding specifications for

BP-1 using several point mutants (pm7, pm51, pm87, and pm102) that have previously been analyzed for interaction with E1B 19K-associated proteins as well as for their ability to inhibit apoptosis. While pm7 and pm102 retained ability to bind to BP-1, substitution of either phenylalanine for serine at position 51 (pm51) or glycine for alanine at position 87 (pm87) resulted in a loss of binding (Fig 19A). The lack of binding with pm51 is consistent with a role of BH3 in the interaction between E1B 19K and BP-1. Although the loss of binding with pm87 might suggest a role in BH1, it should be noted that the glycine residue at position 87 is absolutely conserved within the Bcl-2 family and is located in an integral region adjacent to the hydrophobic cleft which serves as the BH3 binding pocket. Therefore, the pm87 mutant may interfere with the BH3 region and/or result in a highly misfolded protein. Indeed, pm87 is generally defective in binding and at inhibiting apoptosis. Thus, taken together, the mutational analyses indicate that the BH3 and the adjacent N-terminal sequences (possibly BH4) may play a more critical role in the interaction with BP-1. This binding profile overlaps but is distinct from that for Bax and Nbk/Bik and corresponds to a region of E1B 19K that is required for inhibition of apoptosis.

Characterization of BP-1

To determine the size and distribution of *bp-1*, we performed a Northern blot using poly A⁺ RNA prepared from various tissues. Two transcripts were detected at 5 kB and 3 kB which appeared to be ubiquitously expressed, although only low levels were detected in the liver (Fig. 20A). The larger transcript appeared to be

expressed more abundantly than the shorter form. Since the largest fragment of *bp-1* obtained in the two-hybrid screen was only 1.5 kB, we sought to recover full-length cDNAs corresponding to *bp-1*. Using conventional library screening techniques along with database searches we were able to identify both full length transcripts which were named *btf* (*Bcl-2*-associated *transcription factor*). The 3 kB transcript, *btf_s*, was isolated by screening a HeLa λ -cDNA library. The 3'-end of *btf_s* was identical to *bp-1* except that it was missing 147 base pairs (bp) within the predicted coding sequence. While the 5 kB transcript, *btf_L*, could not be recovered from the λ screen, it was identified as a full-length expressed sequence tag (EST) within GenBank (accession # D79986) that was isolated from the human KG-1 cell line. Unlike *btf_s*, *btf_L* did contain the 147 bp region present in *bp-1*. Based on sequence comparisons from GenBank with *btf_s*, we found that the remainder of the coding sequences between *btf_s* and *btf_L* were the same, and the large size difference between the transcripts results from different 3'-untranslated regions (UTRs).

The EST encoding *Btf_L* was utilized to identify the sub-chromosomal location of *btf*. However, it remains to be determined whether *btf_L* and *btf_s* are formed from separate genes or whether they are generated by alternative splicing of the same gene. Using the UniGene collection assessed through National Center for Biotechnology Information (NCBI), we found a 137 bp PCR fragment within the 3'-UTR of the EST (dbSTS entry G20483) that mapped to chromosome 6 between markers D6S292 and D6S1699. These markers correspond to 6q22-23, a locus with a high frequency of deletions in tumors, particularly lymphomas and leukemias (Mitelman,

1995). Thus, the locus of *btf* correlates with a chromosomal region that may contain a tumor suppressor gene.

To test whether *btf* is actually deleted in tumors, we performed a Northern blot using mRNA prepared from several human cancer cell lines. While both transcripts of *btf* were observed in most cell lines tested, they appeared to be less abundant than normal cells and *btf* was not detected in Raji cells which were derived from Burkitt's lymphoma (Fig. 20B). Indeed, Raji cells were the one cell line tested with known 6q deletions. The Northern blot data was therefore consistent with the chromosomal mapping of the gene. Thus, loss of *btf* may actually contribute to tumor formation in these cells.

The primary amino acid sequence encoded by the long and short forms of *btf* are compared in Fig. 21. Btf_L is 918 amino acids and has a predicted molecular mass of 106 kDa, whereas Btf_S is missing 49 amino acids near the carboxyl terminus (amino acids 797-846 of Btf_L) and has a predicted size of 101 kDa. See Figures 21 and 22. The original *bp-1* clone encodes for amino acids 384-918 of Btf_L (Btf ΔN384) which is slightly more than half of the full-length protein (Fig. 22). In general, the 49 amino acid region specific to Btf_L and ΔN384 contains highly charged residues (48%), but the significance of this region is not known. Both Btf_S and Btf_L were able to bind E1B 19K in the yeast two-hybrid assay (data not shown). While the overall protein sequence of Btf is not significantly homologous to other known proteins, searches for conserved motifs using the PROSITE database revealed two nuclear localization sequences (NLS) (Fig. 21) as well as putative DNA-binding domains (Fig. 22). There was 88% homology to the basic zipper (bZIP) DNA-

binding domain between amino acids 110 and 126, and 80% homology to the Myb DNA-binding domain within amino acids 522 and 531 of Btf (Fig. 22).

5 *Btf_s is a nuclear DNA-binding protein that is sequestered by the Bcl-2 family*

 Since *btf_s*, but not *btf_L*, was isolated from the HeLa library, we concentrated our functional assays using the shorter variant. *Btf_s* was cloned into the pcDNA3 vector with a Myc epitope for *in vitro* translation and mammalian expression. The protein sequence of *Btf_s* suggested that it may bind to DNA. To test this hypothesis ³⁵[S]-labeled *in vitro* translated Myc-Btf_s was incubated with native DNA-cellulose. DNA-binding was detected with *Btf_s* but not with E1B 19K used as a negative control (Fig. 23). The strength of the interaction was comparable to a known transcription factor, Msx-1, and was not competed by RNA (data not shown). *In vitro* translated E1B 19K and Bcl-2 (data not shown) did not inhibit the ability of *Btf_s* to bind to DNA suggesting that the binding sites for DNA and E1B 19K/Bcl-2 are in distinct domains within *Btf_s*. Indeed, comparison of the putative DNA-binding domains and the BP-1 (Btf ΔN384) protein which is sufficient for binding to the Bcl-2 family members is consistent with there being two separate domains.

 To confirm the association between *Btf_s* and Bcl-2 family members we performed an *in vitro* binding assay. ³⁵[S]-labeled *in vitro* translated Myc-Btf_s or Myc-Bax, used as a positive control, was combined with E1B 19K, Bcl-2, and Bcl-x_L. Immunoprecipitation with an antibody against Myc revealed that *Btf_s* bound to all three of the anti-apoptotic Bcl-2 family members, but not to

luciferase used a negative control (Fig. 24). However, the strength of these associations were weaker than those with Bax (Fig. 24). These results support the data obtained from the yeast two-hybrid assay.

5 To examine the biological of Btf_s *in vivo*, we attempted to express the tagged protein in HeLa cells. While were unable to obtain stable expression of Btf_s even using an inducible promoter, transient transfection did produce low levels of Btf_s. Although the levels of
10 Btf_s were not high enough to detect by Western blot analysis of whole cell extracts (data not shown) which would allow us to examine whether Btf_s could co-immunoprecipitate with Bcl-2 family members, we were able to visualize Btf_s by immunofluorescence. Its
15 expression was confined to the nucleus and was present in roughly 4% of the cells (Fig 25). This was consistent with the DNA-binding activity, but not with the established localization of Bcl-2-like proteins which are generally found associated with membrane
20 structures, particularly the mitochondria, endoplasmic reticulum, and nuclear envelope. Unlike other Bcl-2 family members, E1B 19K is not normally present in the mitochondria, but rather predominantly localized to the nuclear envelope. However, since the Bcl-2 family can
25 act by sequestering proteins we checked whether E1B 19K, Bcl-2, and Bcl-x_L could alter the subcellular localization of Btf_s. First, E1B 19K and Bcl-2 were co-transfected with Myc-Btf_s. The cells were fixed 24
30 hours post-transfection and then double stained for the Myc epitope present on Btf_s (rhodamine) and either E1B 19K or Bcl-2 (fluorescein). While the percent of Btf_s-positive cells was similar to that when it was transfected by itself, the subcellular localization was

altered. Btf₅ co-localized with E1B 19K and Bcl-2 within the cytoplasm and nuclear periphery in almost all co-expressing cells (Fig. 25). Subsequent experiments were performed to determine if Bcl-x_L could also sequester Btf₅ since these proteins also bound in the yeast two-hybrid assay and *in vitro*. We could not co-stain for Btf₅ and Bcl-x_L with available antibodies, so we developed a stable HeLa cell line expressing Bcl-x_L and stained for Btf₅ using the antibody against the Myc epitope. While control HeLa cells contained 100% nuclear Btf₅ expression, only 7% of the Btf₅-positive cells in the HeLa-Bcl-x_L cell line displayed nuclear Btf₅ staining and almost all of the Btf₅ staining was in the cytoplasm in a pattern similar to Bcl-x_L (Fig. 26). The few cells expressing nuclear Btf₅ may be accounted for by the variable levels of Bcl-x_L observed in this cell line and/or an incomplete ability of Bcl-x_L to sequester Btf₅. Nevertheless, it is in clear contrast to the parental HeLa cells where the expression of Btf₅ was completely nuclear. These results suggest that E1B 19K, Bcl-2, and Bcl-x_L can alter the localization, and therefore apparently, the function, of Btf₅.

Btf₅ represses transcription which is inhibited by the Bcl-2 family

Since Btf₅ was shown to bind DNA, we checked whether it could modulate transcription. We first performed a one-hybrid assay in yeast by generating a fusion protein of Btf₅ with the GAL-4 DNA-binding domain in the pGBT9 vector. If Btf₅ contained a *trans*-activation domain one would expect that the adjacent activation and DNA-binding domains would lead to GAL-4-inducible phenotypes in yeast. Growth was not detected in the absence of

histidine suggesting that Btf₈ does not contain a transcriptional activation domain (data not shown). However, it remains possible that Btf₈ requires mammalian co-factors in order to activate transcription or that Btf₈ represses, rather than activates, transcription.

To address these issues, Btf₈ was cloned into a mammalian expression vector (pml) containing the GAL-4 DNA binding domain. Transcriptional activity was monitored using a reporter construct containing GAL-4 promoter sites. Transient expression of pml-Btf₈ with the luciferase reporter led to nearly a 10-fold decrease in transcription compared to an empty pml vector control (Fig. 27). This effect is comparable to the trans-repressive activity of other proteins, such as p53 and Msx-1. Two deletion mutants of Btf₈, ΔN522 (amino acids 522-918 without residues 797-846 and ΔC210 (amino acids 1-210) were also cloned into pml to determine the general regions that may contribute to transcriptional repression (Fig. 22). Both deletion mutants of Btf₈ were detectably expressed in HeLa cells to levels comparable with wild-type Btf₈ (data not shown). While Btf₈ΔN522 was not able to repress transcription, the ΔC210 mutant was sufficient for repression and nearly as potent as full-length Btf₈ (Fig. 27). Interestingly, this fragment which contains the bZIP homology segment is also rich in serine and glycine residues, a feature that is present in other transcriptional repressors.

Since the Bcl-2-like proteins were capable of sequestering Btf₈ in the cytoplasm, we hypothesized that they may also block the ability of Btf₈ to repress transcription. To test this possibility we co-transfected ElB 19K, Bcl-2, and Bcl-x_L with pml-Btf₈ and the luciferase reporter construct. In the control

samples, none of the Bcl-2 family of proteins had a significant effect on transcription of the luciferase reporter (Fig. 27). However, transfection of any of the three Bcl-2 family members (E1B 19K, Bcl-2, and Bcl-x_L), but not the empty pcDNA3 vector, abrogated Btf₂-mediated transcriptional repression. These data suggest that E1B 19K, Bcl-2, and Bcl-x_L inhibit Btf₂ trans-repression, probably by binding to and sequestering Btf₂ in the cytoplasm.

Sustained expression of Btf₂ inhibits transformation

Transfection of the adenovirus E1A gene along with E1B 19K or bcl-2 in primary BRK cells induces transformation as a consequence of dual proliferative and anti-apoptotic signaling. Expression of E1B 19K/Bcl-2 binding proteins, such as Bax and Nbk/Bik, antagonizes the anti-apoptotic signal and causes a reduction in foci formation. To determine if Btf₂ could also antagonize the ability of E1B 19K to transform primary cells, we transfected E1A and E1B 19K, with and without Myc-Btf₂, into primary BRK cells. Btf₂ caused a 64% reduction in foci formation mediated by E1A and E1B 19K (Fig. 28A). This suggests that interaction between Btf₂ and E1B 19K inhibits 19K function *in vivo* and/or Btf₂ is capable of suppressing transformation independent of 19K. The reduction in foci formation by Btf₂ and E1A compared to E1A alone suggested the later is possible (Fig. 28A). However, we tested whether Btf₂ could inhibit another transforming signal, E1A with p53DD. p53DD is a p53 deletion mutant that contains the C-terminal oligomerization domain and functions in a dominant-negative fashion to inhibit p53-mediated apoptosis and growth arrest. Since p53DD blocks both of

these processes, it produces a very potent transforming signal. Here we show that expression of Btf₅ was capable of repressing foci formation by E1A and p53DD by about 60%, suggesting that btf₅ may act as a general suppressor of transformation (Fig. 28B).

Btf₅ functions to induce apoptosis

To further characterize the transformation suppressing activity of Btf₅, we tested the effect of its expression on apoptosis and cell cycle progression. Attempts to generate stable BRK or HeLa cell lines expressing Btf₅ failed, indicating that sustained Btf₅ expression is incompatible with either cell proliferation or viability. To monitor Btf₅ expression in transient assays, the cDNA was cloned into the pIRES-EGFP vector which provides co-expression of Btf₅ and the enhanced green fluorescent protein (EGFP) marker. HeLa cells were transiently transfected and the cells were harvested 48 and 72 hours post-transfection. The cell cycle kinetics were analyzed following propidium iodide staining by FACS analysis. Since EGFP staining with Btf₅ was not detectable until 48 hours post-transfection, we were unable to characterize the cell cycle characteristics at the earlier time points. The levels of EGFP expression and the cell cycle kinetics at 48 and 72 hours are shown in Table 1. At 48 hours post-transfection there were only 19.5% EGFP-positive cells in the presence of Btf₅, whereas there were 61.2% positive cells in the control pIRES-EGFP empty vector. The FACS analysis demonstrated that Btf₅ led to an increase in percent of subG_{0/1} cells from 3.4% to 15.8% indicating an increase in cell death. No other obvious changes in cell cycle parameters were observed at this

time point.

To test whether E1B 19K could inhibit this cell death, we co-transfected pIRES-EGFP-Btf_s with pCMV-E1B 19K. E1B 19K inhibited Btf_s-mediated cell death at 48 hours as indicated by an increase in the number of EGFP expressing cells from 19.5% to 34.3% and fewer subG_{0/1} cells (15.8% to 10.1%) (Table 1). Thus, E1B 19K expression abrogated cell death induced by Btf_s. At 72 post-transfection the amount of cell death triggered by Btf_s increased and could no longer be inhibited by E1B

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TABLE 1. Overexpression of Btfa induces cell death^a

Vector	48 h posttransfection					72 h posttransfection				
	% EGFP positiv e	% sub- G ₀ /1	% G ₀ /1	% S	% G ₂ /M	%EGFP positi ve	% sub- G ₀ /1	% S	% G ₂ /M	% G ₂ /M
pIRES-EGFP + pCDNA3	61.2	3.4	42.3	20.0	33.7	45.7	5.6	32.5	20.9	40.6
pIRES-EGFP + pCMV-E1B 19K	72.0	1.5	54.2	21.0	23.6	59.0	8.8	55.4	20.8	14.8
pIRES-EGFP-Btfa + pCDNA3	19.5	15.8	36.7	21.7	25.6	13.2	29.1	33.2	20.6	16.7
pIRES-EGFP-Btfa + pCMV-E1B 19K	34.3	10.1	41.8	17.8	30.0	15.4	36.9	36.0	13.3	13.6

^a HeLa cells were electroporated with 20 µg of pIRES-EGFP-Btfa, or empty vector in combination with 10 µg of pCMV-E1B 19K or empty pCDNA3 vector. The cells were harvested 48 and 72 h posttransfection, fixed in 2% paraformaldehyde, and stained with PBS containing 1 µg of propidium iodide per ml. 250 µg of FACS A per ml. and 0.1% Tween 20. The cells were incubated for at least 30 min at room temperature and then analyzed by FACS. The results show the percentage of EGFP-positive cells and the cell cycle kinetics for the EGFP-positive population.

19K. In the presence of Btf_s there were only 13.2% EGFP positive-cells, 29.1% of which were represented in the subG_{0/1} peak (Fig. 29). The 72 hour time point also revealed a decrease in the G₂/M peak with Btf_s (16.7%) compared to the control vector (40.6%). The change in G₂/M by Btf_s may be an indication that cells are exiting from these phases of the cell cycle to go into apoptosis.

Interestingly, we also observed a decrease in the G₂/M peak with E1B 19K in the absence of Btf_s. At 72 hours post-transfection there was a change in the number of cells in G₂/M from 40.6% with the control vector to 14.8% in E1B 19K transfected cells. Changes in cell cycle progression by the anti-apoptotic Bcl-2 family members have been observed previously, such that generally they produce a decrease in cell cycle progression. While others have shown that Bcl-2 causes an increase in G_{0/1}, here we show that E1B 19K causes an decrease in G₂/M. Thus, these results may reflect a difference in the mechanism between the cell cycle regulation by E1B 19K and other related proteins.

Cell death has been classified as either necrosis or apoptosis, where necrosis is considered a passive process of cell death associated with trauma, and apoptosis is a genetically programmed active response leading to cell suicide. Unlike necrosis, apoptosis involves morphological changes such as chromatin condensation, DNA fragmentation, and cytoplasmic blebbing. To test whether the increase in subG_{0/1} cells by Btf_s was due to apoptosis, we examined the nuclei of EGFP-positive cells with Hoechst dye 72 hours post-transfection. Chromatin condensation observed by this approach would be one indication of apoptosis. HeLa

cells transfected with the empty pIRES-EGFP vector showed EGFP stained cells with normal nuclei. However, in cells transfected with pIRES-EGFP-Btf_s, the EGFP-positive cells had condensed chromatin staining, indicating that they were dying by apoptosis (Fig. 30 Taken together, these results suggest that Btf_s can function to promote apoptosis which may account for its ability to suppress transformation. E1B 19K can inhibit Btf_s-induced apoptosis, albeit incompletely in some assays. None the less, this suggests that the Bcl-2 family affects apoptosis through modulation of transcription.

DISCUSSION

The Bcl-2 family regulates apoptosis through multiple mechanisms. For example, these proteins can function at the mitochondria by possibly forming channels that regulate mitochondrial membrane potential and cytochrome C release. Alternatively, the Bcl-2 family may directly regulate caspase activation through interactions with Ced-4-like proteins. We describe a novel E1B 19K-interacting protein, Btf, isolated through a two-hybrid screen that regulates an alternative pathway to control apoptosis. Two transcripts corresponding to *btf* were identified, *btf_s* and *btf_L*. Both appeared to be widely expressed, but were deleted in some tumors. Here, we described the function and biological significance of the protein product generated from the shorter form, Btf_s, which differs from Btf_L in just a 49 amino acids in the C-terminal region.

Double staining experiments were able to show the co-localization of Btf_s with E1B 19K, Bcl-2 and Bcl-x_L, thereby supporting the interactions observed in the

yeast two-hybrid assay as well as by *in vitro* co-immunoprecipitation. While transiently expressed Btf_s was nuclear and could be sequestered into the cytoplasm by the anti-apoptotic Bcl-2 family members, the localization of endogenous Btf_s remains to be determined. It is entirely possible that in normal, non-apoptotic cells Btf_s is predominantly expressed in the cytoplasm. In contrast, in dying cells, where there is a higher percentage of pro-apoptotic versus anti-apoptotic Bcl-2 family members, Btf_s may translocate to the nucleus and potentiate apoptosis.

Btf_s was able to bind DNA *in vitro*, and in reporter assays Btf_s repressed transcription. The *trans*-repressive activity was inhibited by ElB 19K, Bcl-2 and Bcl-x_L, correlating with their cytoplasmic sequestration potentials. While it remains possible that part of this phenomenon is a result of Btf_s-induced apoptosis, the amount of cell death observed at 24 hours would not account for the 10-fold reduction in transcriptional activity. The evidence that Btf_s is a transcription repressor was further supported by the identification of the ΔC210 deletion mutant that was sufficient for repressing transcription. The N-terminal fragment of Btf_s is rich in glycine and serine, a feature that is common to transcriptional repressors. Taken together, these data provide evidence for a novel *trans*-repressive protein that triggers apoptosis which is sequestered and inhibited by Bcl-2 family members.

One of the features of the Bcl-2 family exemplified by the Btf_s interaction is their ability to sequester other cellular proteins from their normal subcellular localization. This process enables ElB 19K and Bcl-2 to inhibit apoptosis using more than one cellular pathway.

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The best characterized example concerns the association between pro-apoptotic and anti-apoptotic Bcl-2 family members. While these interactions have been known for several years, recent studies of E1B 19K and Bax have demonstrated that these family members can alter each other's sub-cellular localization. Bax is normally stimulated to go to the mitochondria during apoptosis and causes a loss in mitochondrial membrane potential. However, over-expression of E1B 19K causes Bax to be sequestered to the nuclear periphery where E1B 19K is localized. This process most likely blocks the ability of Bax to disrupt mitochondrial function. The Bcl-2 family also associates with several unrelated proteins that may contribute to apoptosis regulation. For example, Bcl-2 associates with and targets the serine/threonine kinase Raf-1 to the mitochondria. Over-expression of Bcl-2 and Raf-1 cooperate to inhibit apoptosis. Another example that is now emerging is the association between the anti-apoptotic Bcl-2 family members with *C. elegans*, Ced-4, and with its mammalian homologue Apaf-1. These interactions directly block the activation of downstream caspases. Thus far, the *C. elegans* Bcl-2 homologue, Ced-9, as well as Bcl-x_L and E1B 19K have been shown to bind to Ced-4, and at least Ced-9 and E1B 19K can redistribute Ced-4 from the cytosol to cytoplasmic membranes. Bcl-x_L also interacts with Apaf-1 and therefore it will be interesting to determine whether Bcl-x_L can alter the localization of Apaf-1. In perhaps an analogous scenario, E1B 19K also sequesters the death-promoting protein FADD, an upstream component of Fas- and TNF- α -mediated death signaling pathway. Overexpressed FADD becomes multimerized and produces filaments throughout the cell. E1B 19K disrupts the

FADD filaments causing FADD to relocate with regions normally associated with 19K and inhibits FADD-dependent apoptosis. Here, we show that the Bcl-2 family members also sequester a nuclear transcription factor and that this may also play a role in apoptosis. The minimal region for ElB 19K required for interaction with Btf at the amino terminus (amino acids 1-36) appears to be distinct from the interaction regions involved in binding to other pro-apoptotic proteins (Bax, Ced-4, Nbk/Bik). This amino terminal region of ElB 19K may correspond to BH4 of Bcl-2 and Bcl-x_L, although this remains to be tested directly. Together these studies suggest that ElB 19K and possibly other Bcl-2 family members, act as binding proteins for a number of apoptosis regulators which could contribute to their widespread role as apoptosis inhibitors.

Transcriptional regulation often plays a critical role during apoptosis by either activating or repressing genes encoding basic apoptotic components. Indeed, inhibition of RNA and protein synthesis block apoptosis induced by a number of circumstances, including growth factor deprivation and following treatment with some chemotherapeutic drugs. In contrast, others have shown that these inhibitors can actually promote cell death, suggesting that loss of a short-lived survival factor can also lead to apoptosis. A number of transcription factors have been identified that may serve as positive or negative regulators of apoptosis. For example, the NF- κ B transcription factor plays an important role in blocking apoptosis triggered by TNF- α although the target genes for NF- κ B are not yet known. However, in other situations, NF- κ B activation has been associated with induction of apoptosis. Furthermore, Bcl-2 and ElB

19K represses NF- κ B activity providing another mechanism for Bcl-2 family members to control apoptosis.

Another transcription factor that may be regulated by the Bcl-2 family during apoptosis is p53. The p53 tumor suppressor protein is required for apoptosis during ionizing radiation and chemotherapeutic drugs, as well as transforming oncogenes such as *c-myc* and *E1A*. While p53 may have multiple functions, the transcriptional activity of p53 is clearly critical for the regulation of cell death in some situations but not all. However, it is still unclear whether p53-mediated apoptosis requires its transcriptional activation or repression properties, or perhaps both. On the one hand, p53 *trans*-activates both *bax* and *fas*, both of which are bonafide inducers of apoptosis. In contrast, several p53-repressible genes have been identified which may potentially contribute toward apoptosis, including *bcl-2*, *MAP4*, *interleukin-6*, *c-fos*, and *c-myc*. Expression of Bcl-2 and E1B 19K alleviate the transcriptional repression activity of p53 providing a mechanism for Bcl-2 family regulation of apoptosis. Thus, Bcl-2 family members can influence apoptosis by modulating the transcriptional activity of both NF-B and p53. Inhibition of Btf₂-mediated transcriptional repression and apoptosis by the Bcl-2 family establishes another connection between these apoptosis regulators and modulation of transcription. It will certainly be interesting to determine whether Btf₂ can influence the activities of other transcription factors related to apoptosis, such as NF- κ B and p53.

We would predict that Btf₂ may repress the transcription of survival genes. Based on previous observations of regulation of gene expression by E1B

19K, one possible target may be the p53 inhibitor, Mdm-2. We have previously observed that stable expression of E1B 19K or Bcl-2 leads to an increase in Mdm-2 mRNA and protein levels. Such a mechanism could also account for the ability of E1B 19K to derepress the transcriptional repression mediated by p53 and E1A. While it will be worthwhile to determine whether Btf₂ could affect p53 modulation of these targets, the fact that Btf₂ was capable of inhibiting transformation by p53DD suggests that another target also exists that is independent of p53. We also have not been able to detect binding between Btf₂ and p53 (data not shown), although it is still plausible that Btf₂ associates with p53-dependent co-activators such as p300. Since the putative DNA-binding sites within Btf₂ contains homology to bZIP and Myb, it is conceivable that they share similar targets. Interestingly, correlations exist between both of these transcription factor families with apoptotic regulation. For example, application of functional blocking antibodies against the AP-1 proteins, Fos and Jun, or transfection with dominant-interfering Jun, inhibits apoptosis in neuronal cells following growth factor withdrawal. However, relevant target genes that contribute toward AP-1-mediated apoptosis have yet to be identified. The Myb family of transcriptional regulators inhibit apoptosis, and regulate the transcriptional activation of bcl-2. This may therefore provide a feedback loop between Btf₂ and Bcl-2. Future studies will be performed to determine whether Btf₂ regulates bcl-2, or other genes involved in apoptosis. The link between Btf₂ and the Bcl-2 family may provide an alternative mechanism by which the Bcl-2 family is able to regulate cell survival.

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While certain of the preferred embodiments of the present invention have been described and specifically exemplified above, it is not intended that the invention be limited to such embodiments. Various modifications may be made thereto without departing from the scope and spirit of the present invention, as set forth in the following claims.

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What is claimed is:

1. A recombinant cell line for assessing therapeutic agents that regulate apoptosis, comprising:

- 5 a) a first plasmid expressing a p300 responsive promoter operably linked to a first reporter gene;
- b) a second plasmid expressing a non p300 responsive promoter operably linked to a second reporter gene; and
- 10 c) a third plasmid expressing a selectable marker gene.

2. The cell line of claim 1, said cell line being stably transfected with an additional plasmid encoding wild -type p300 to augment endogenously expressed p300 protein levels.

15

3. A screening method for determining if a therapeutic reagent inhibits p300 activity thereby inducing apoptosis, comprising:

20

- a) contacting recombinant cells with said therapeutic agent, said cells containing
- i) a first plasmid expressing a p300 responsive promoter operably linked to a first reporter gene;
- 25

ii) a second plasmid expressing a non p300 responsive promoter operably linked to a second reporter gene; and

iii) a third plasmid expressing a selectable marker gene;

30

b) assessing cells for repression of the p300 responsive reporter gene by said therapeutic reagent; and

c) assessing cells for repression of the non-p300 responsive reporter gene by said therapeutic reagent, repression in step b) and not step c) indicating that the compound inhibits p300 transactivation and thereby induces apoptosis.

4. A recombinant cell line for assessing therapeutic agents that regulate apoptosis, comprising:

- a) a first plasmid expressing Btf; and
- b) a second plasmid expressing a Bcl-2.

5. A screening method for determining if a therapeutic reagent inhibits Bcl-2 binding to Btf thereby inducing apoptosis, comprising:

- a) contacting recombinant cells with said therapeutic agent, said cells containing
 - i) a first plasmid expressing btf;
 - ii) a second plasmid expressing Bcl-2;
- b) assessing cells for disruption of Bcl-2 binding to btf by said therapeutic reagent; and
- c) determining whether said disruption induces apoptosis in said recombinant cells.

6. A method for detecting the presence of btf encoding nucleic acids in a tumor sample, comprising:

- a) obtaining a tumor sample suspected of containing a deletion of btf encoding nucleic acids;
- b) isolating DNA from said sample;
- c) contacting said DNA with a nucleic acid comprising a sequence of SEQ ID NO: 1 under conditions whereby said DNA will hybridize with SEQ ID NO:1, if said sample contains btf hybridizable nucleic acids; and
- d) detecting said hybridization if any.

7. A method for detecting Btf protein in a tumor sample, comprising:

- a) providing a tumor sample;
- b) contacting said sample with an antibody immunologically specific for Btf under conditions whereby said antibody will bind said Btf if present; and
- c) detecting said antibody-Btf complex in said sample.

8. A method for introducing btf encoding nucleic acids into a host cell, comprising:

- a) providing a btf encoding nucleic acid in a suitable expression vector, and
- b) delivering said btf encoding nucleic acid to said cell in an effective amount to produce Btf protein.

Fig. 1B

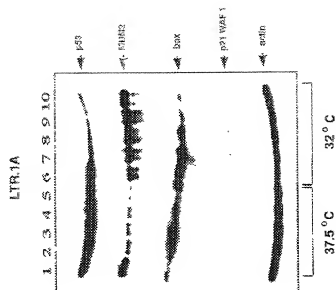
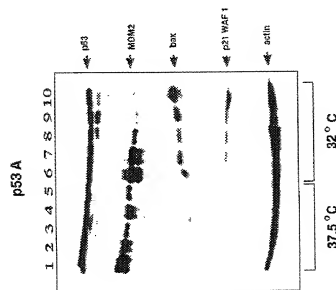
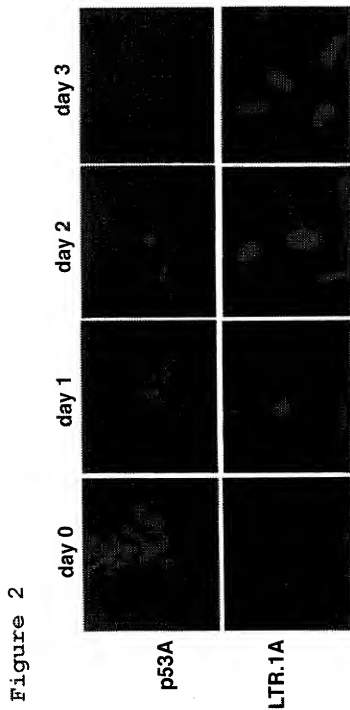


Fig. 1A



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Figure 3A

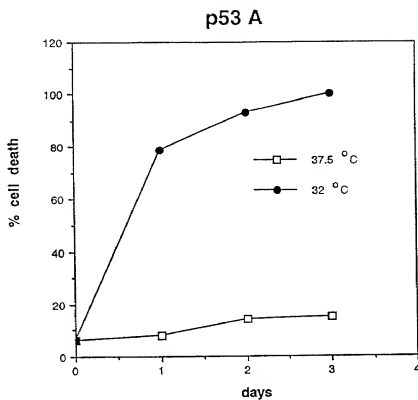
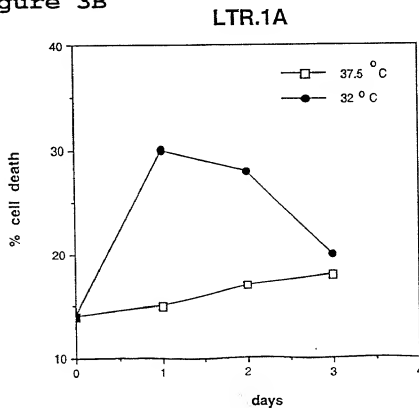


Figure 3B



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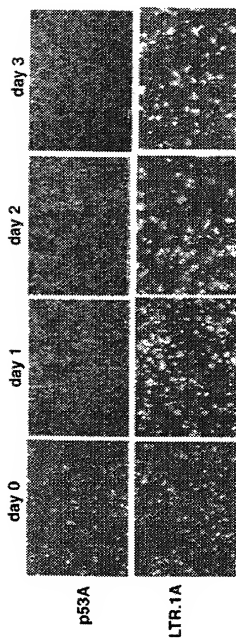


Figure 4

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Fig. 5A

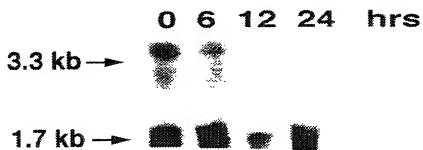
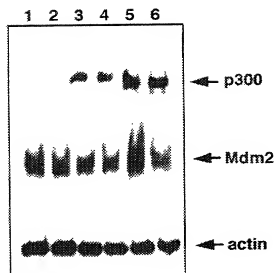


Fig. 5B



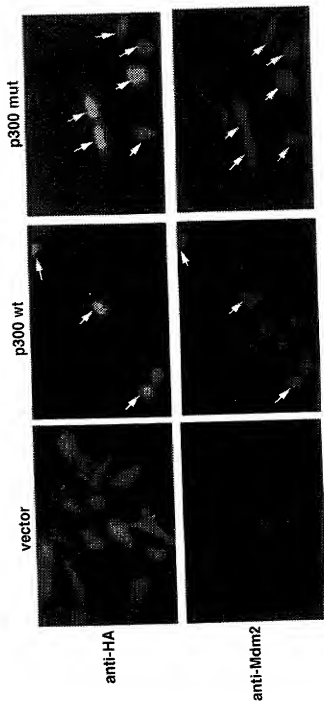
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Figure 6



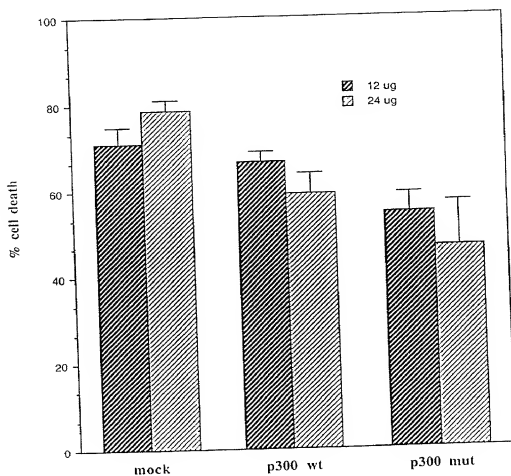
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Figure 7



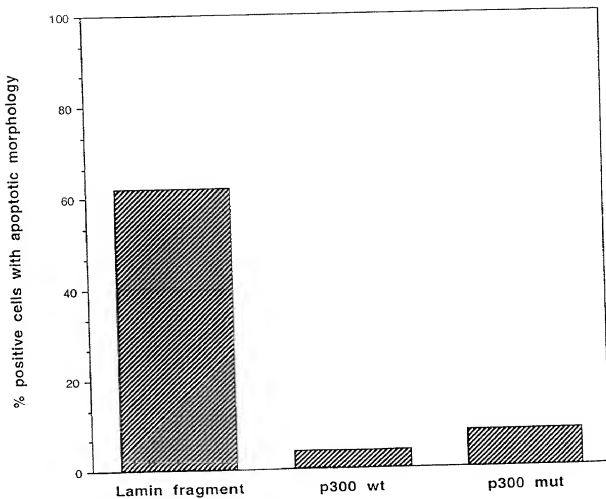
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Figure 8



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Figure 9

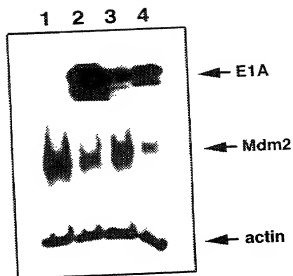


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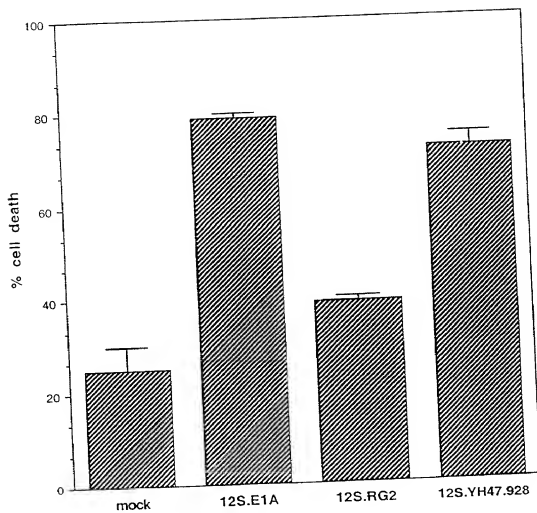
PCT/US99/09793

Figure 10



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Figure 11



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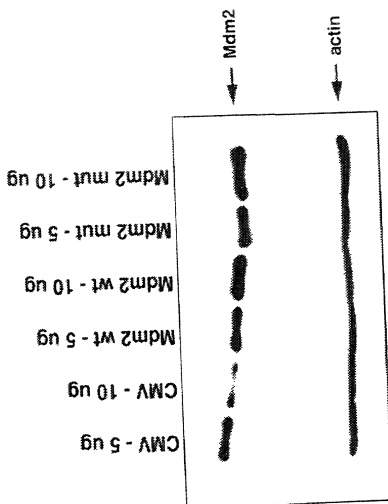
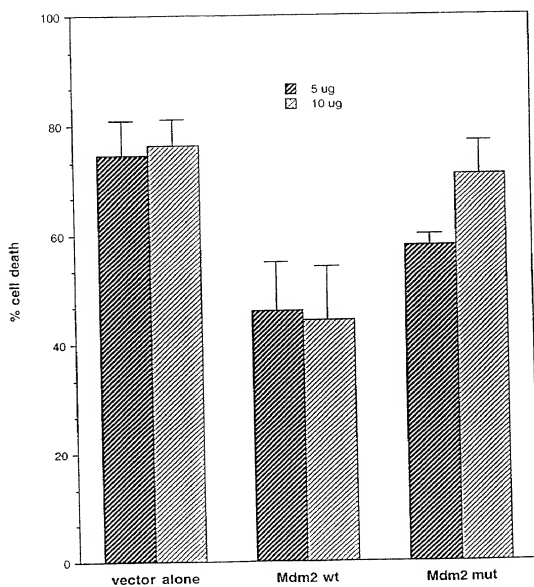


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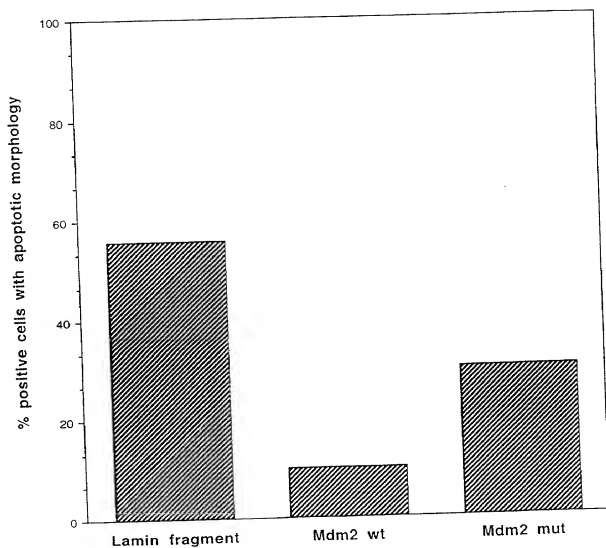
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Figure 13



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Figure 14



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Fig. 15C

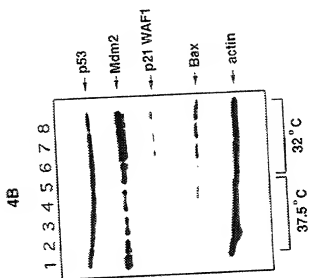


Fig. 15B

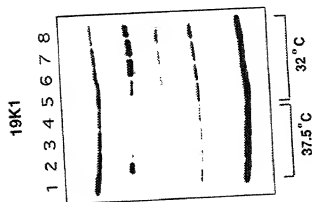
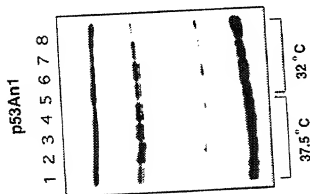
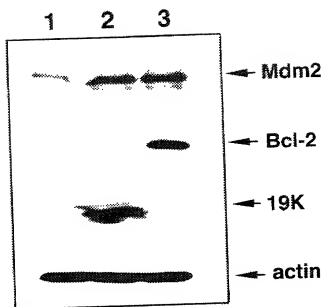


Fig. 15A



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Figure 16



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Fig. 17A

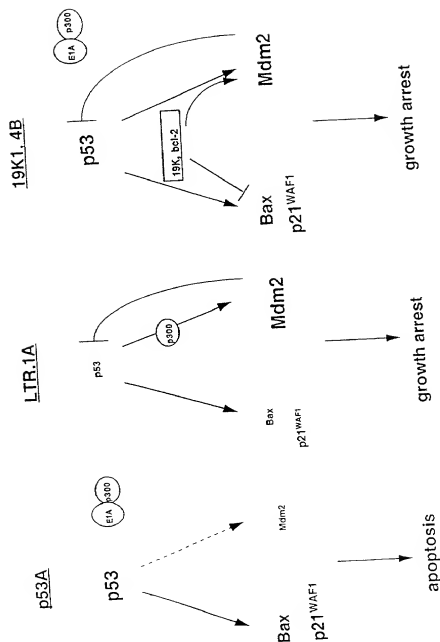


Fig. 17B



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Figure 18



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Fig. 19A

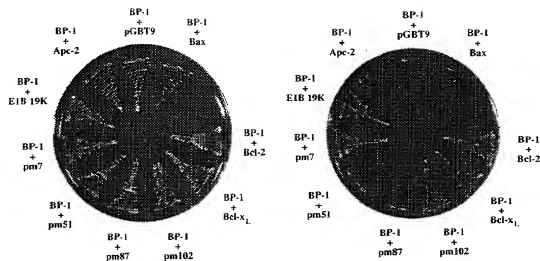


Fig. 19B

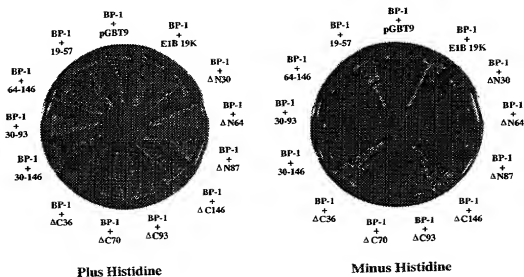
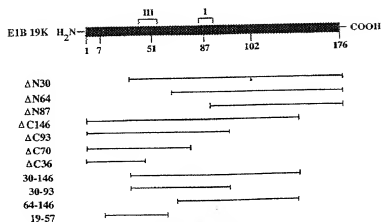


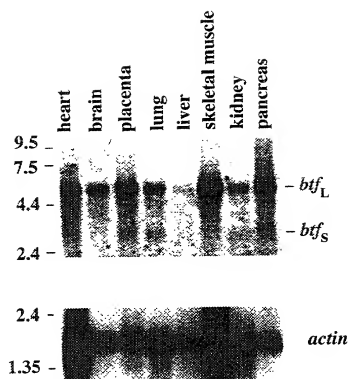
Fig. 19C



SUBSTITUTE SHEET (RULE 26)

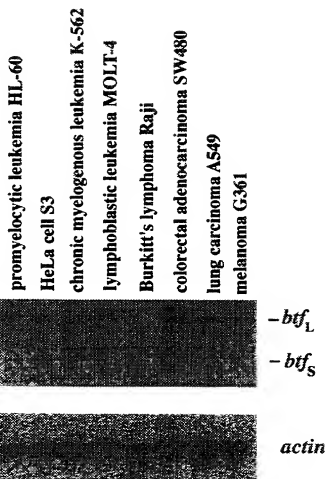
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Fig. 20A



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Fig. 20B



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Figure 21

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Figure 22

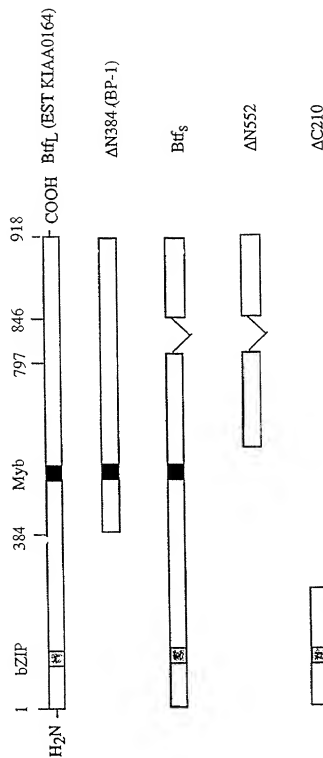
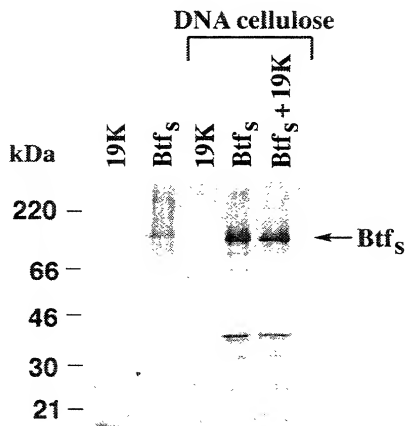


Figure 23



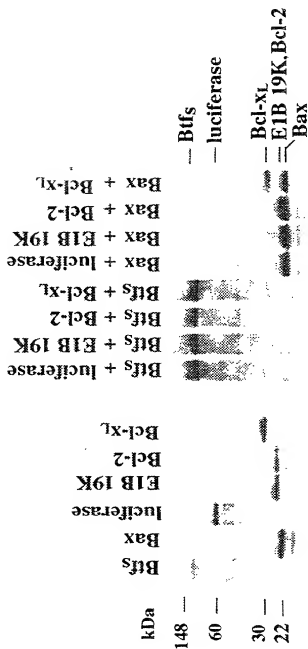
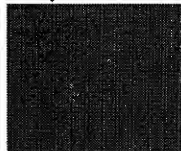
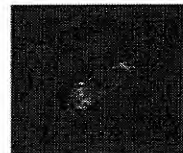


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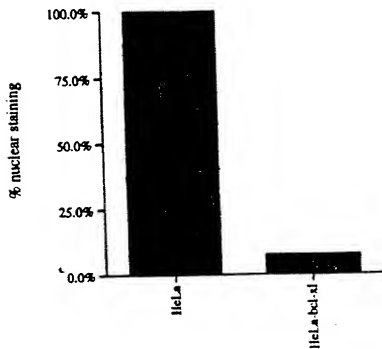
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Figure 25

pcDNA3
anti-MycBtf_s
anti-MycBtf_s + E1B 19K
anti-MycBtf_s + E1B 19K
anti-E1B 19KBtf_s + Bcl-2
anti-MycBtf_s + Bcl-2
anti-Bcl-2

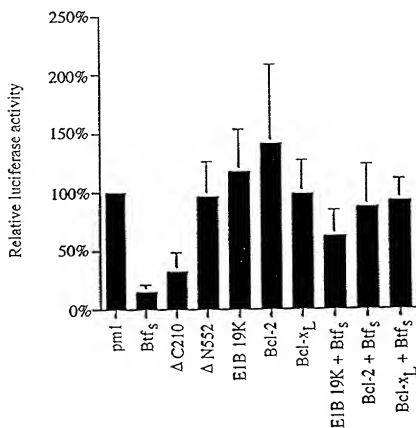
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Figure 26



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Figure 27



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Figure 28A

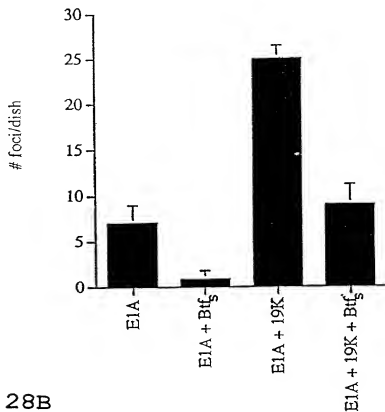
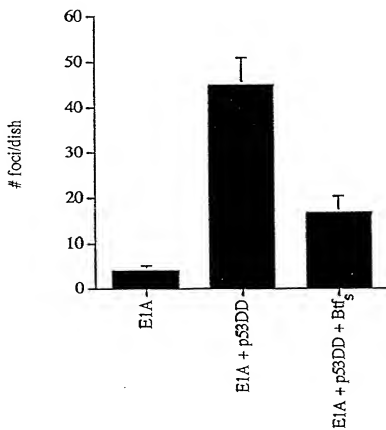


Figure 28B



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Figure 29

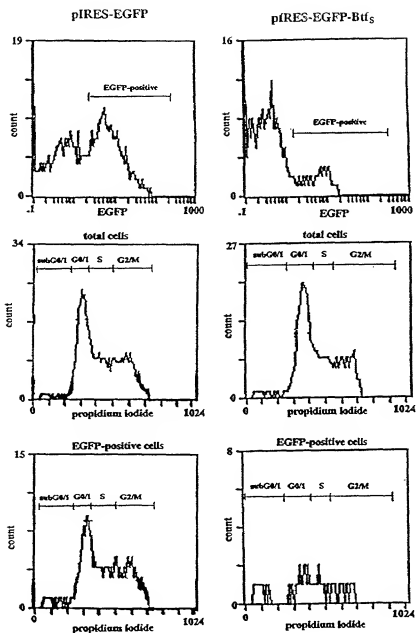
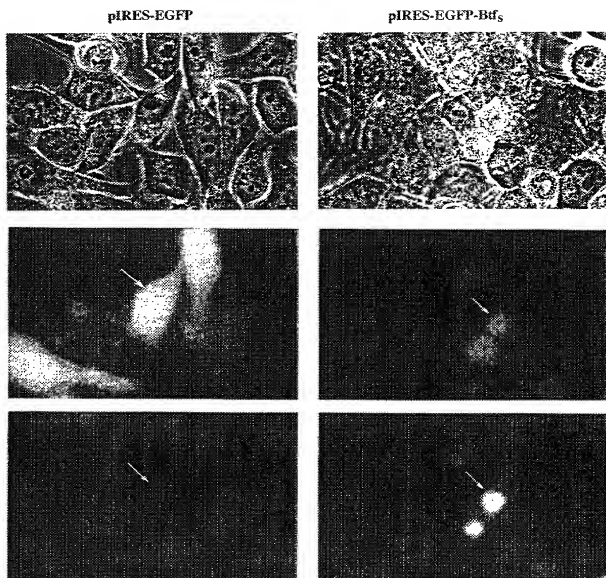


Figure 30



DECLARATION, POWER OF ATTORNEY AND POWER TO INSPECT

As a below named inventor, I hereby declare:

that my residence, post office address and citizenship are as stated below next to my name;

that I verily believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural inventors are named below) of the invention entitled: **RECOMBINANT CELL LINE AND SCREENING METHOD FOR IDENTIFYING AGENTS WHICH REGULATE APOPTOSIS AND TUMOR SUPPRESSION**

the specification of which (check one(s) applicable)

☒ was filed May 6, 1999 as International Application No. PCT/US99/09793, on which U.S. Patent

Application No. 09/674,876 is based.

and was amended by Amendment filed _____ (if applicable); [or],

is attached to this Declaration, Power of Attorney and Power to Inspect;

that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above; and that I acknowledge my duty to disclose information which is material to the examination of this application in accordance with Rule 56 (a) [37 C.F.R. §1.56(a)].

CLAIM UNDER 35 USC §119(e): I hereby claim the benefit under 35 USC §119(e) of any United States provisional applications listed below:

<u>Provisional Application No.</u>	<u>Filing Date</u> <u>Day/Mo/Yr</u>
60/084,664	7 May 1998
60/081,391	1 July 1998
60/092,871	15 July 1998
60/107,689	9 November 1998

POWER OF ATTORNEY: As inventor, I hereby appoint **DANN, DORFMAN, HERRELL AND SKILLMAN, P.C.** of Philadelphia, PA, and the following individual(s) as my attorneys or agents with full power of substitution to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith: **Kathleen D. Rigaut, Ph.D., J.D. Reg. No. 43,647 and Patrick J. Hagan, Reg. No. 27,643**

POWER TO INSPECT: I hereby give **DANN, DORFMAN, HERRELL AND SKILLMAN, P.C.** of Philadelphia, PA or its duly accredited representatives power to inspect and obtain copies of the papers on file relating to this application.

SEND CORRESPONDENCE TO: CUSTOMER NUMBER 000110.

DIRECT INQUIRIES TO: Telephone: (215) 563-4100 **Kathleen D. Rigaut, Ph.D., J.D.**
Facsimile: (215) 563-4044

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

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CK 4/21/0

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SEQUENCE LISTING

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White, Eileen
Kasof, Gary
Goyal, Lakshmi
- <120> Methods and compositions for regulation
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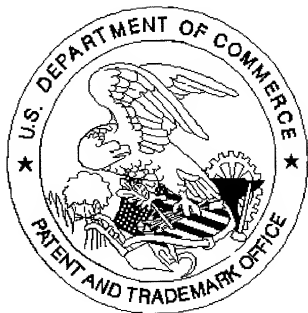
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 Gly Phe Ala Gly Val Ser Arg Pro Arg Gly Thr Phe Phe Arg Ile Arg
 785 790 795 800
 Gly Arg Gly Arg Ala Arg Gly Val Phe Ala Gly Thr Asn Thr Gly Pro
 805 810 815
 Asn Asn Ser Asn Thr Thr Phe Gln Lys Arg Pro Lys Glu Glu Glu Trp
 820 825 830
 Asp Pro Glu Tyr Thr Pro Lys Ser Lys Lys Tyr Phe Leu His Asp Asp

	835		840		845
Arg	Asp	Gly	Val	Asp	Tyr
850				855	
Thr	Phe	Gln	Arg	Gly	Arg
865				870	
Ser	Pro	Lys	Trp	Thr	His
				885	
Asp	Glu	Glu	Glu	Thr	Met
				900	
Lys	Glu	Glu	Lys	Glu	
				915	

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